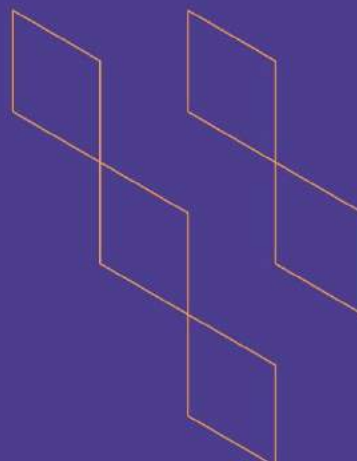




T-104
2022

Course Specification



Course Title: Food safety
Course Code: FSNU313
Program: Food Science and Human Nutrition
Department: Food Science and Human Nutrition
College: Agriculture and Veterinary Medicine
Institution: Qassim University
Version: <i>Course Specification Version Number</i>
Last Revision Date: <i>Pick Revision Date.</i>



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A. General information about the course:

Course Identification	
1. Credit hours:	2 (1+1)
2. Course type	
a.	University <input type="checkbox"/> College <input checked="" type="checkbox"/> Department <input type="checkbox"/> Track <input type="checkbox"/> Others <input type="checkbox"/>
b.	Required <input type="checkbox"/> Elective <input checked="" type="checkbox"/>
3. Level/year at which this course is offered:	College elective
4. Course general Description Introduction of different food safety hazards e.g. chemical, physical, and biological including food pathogenic diseases in food production – pre requisite programs for implementation of food safety systems in food establishment – HACCP and FSMS ISO 22000.	
5. Pre-requirements for this course (if any): BCH301	
6. Co- requirements for this course (if any): Non	
7. Course Main Objective(s) <ul style="list-style-type: none"> • Identify the different hazards affected food safety in the different stages of food production. • Illustrate the basic knowledge of the elements of food safety systems and evaluation of prerequisite programs necessary to implement food safety systems in food establishments. • Gain proficiency in dealing with problems related to food safety and how to solve them. • Get on the sources and compile information relating to food safety systems. 	

1. Teaching mode (mark all that apply)

No	Mode of Instruction	Contact Hours	Percentage
1.	Traditional classroom	15x3 (45)	100%
2.	E-learning		
3.	Hybrid <ul style="list-style-type: none"> • Traditional classroom • E-learning 		
4.	Distance learning		



2. Contact Hours (based on the academic semester)

No	Activity	Contact Hours
1.	Lectures	45
2.	Laboratory/Studio	
3.	Field	
4.	Tutorial	
5.	Others (specify)	
	Total	45

Course Learning Outcomes (CLOs), Teaching Strategies and Assessment Methods

Code	Course Learning Outcomes	Code of CLOs aligned with program	Teaching Strategies	Assessment Methods
1.0	Knowledge and understanding			
1.1				
1.2	Student memorizes the principles of food safety systems such as HACCP and ISO 22000-2005, with understand their prerequisite programs e. g. GMP and GHP.	K2	Lecture, Research activities, Whole group and small group discussion, presentation	Pe, L, F, Hw, Pr, It, Rr, Wt, O
2.0	Skills			
2.1				
2.2				
...				
3.0	Values, autonomy, and responsibility			
3.1	Student illustrates and audits the prerequisite programs and food safety systems e. g. HACCP.	V1	Lecture, Research activities, Whole group and small group discussion, presentation.	Pe, L, F, Hw, Pr, It, Rr, Wt, O
3.2				
...				



C. Course Content

No	List of Topics	Contact Hours
1.	Lecture Topic	
2.	Introduction of food safety and its definitions	1
3.	Local, regional and international bodies concerned with food safety systems and their application in food establishments.	1
4	Different hazards and their sources of contamination affected the safety of food products.	1
5	Different hazards and their sources of contamination affected the safety of food products.	1
6	Risk assessment of different hazards contaminated foods.	1
7	Prerequisite programs of food safety system such as GMP/GHP; SSOP.	1
8	Prerequisite programs of food safety system such as GMP/GHP; SSOP.	1
9	Sanitary Inspection in food establishments.	1
10	Hazard Analysis Critical Control Point (HACCP) system.	1
11	Hazard Analysis Critical Control Point (HACCP) system.	1
12	Hazard Analysis Critical Control Point (HACCP) system.	1
13	Food safety management system (FSMS).	1
14	Food safety management system (FSMS).	1
15	Other food safety systems.	1
16	Auditing food safety system applied in food establishments.	1
1	Practical topic	
2	Introduction to the practical section of the course.	2
3	Risk assessment of different hazards.	2
4	Risk assessment of different hazards.	2
5	Assessment the prerequisite programs of food safety system.	2
6	Assessment the prerequisite programs of food safety system.	2
7	Assessment the prerequisite programs of food safety system.	2
8	Design and Application of food safety systems in food establishments.	2
9	Design and Application of food safety systems in food establishments.	2
10	Design of HACCP plans for different food products.	2
11	Design of HACCP plans for different food products.	2
12	Design of HACCP plans for different food products.	2
13	Design food safety management system in food establishment.	2
14	Design food safety management system in food establishment.	2
15	Auditing of food safety systems.	2
16	Auditing of food safety systems.	2
Total		45



D. Students Assessment Activities

No	Assessment Activities *	Assessment timing (in week no)	Percentage of Total Assessment Score
1.	Periodical exam	5	20%
2.	Practical exam	15	20%
3.	Final exam	End of semester	50%
4.	Periodical activities and home working	5, 7, 8, 10	5%
5	Oral exam	15	5%

*Assessment Activities (i.e., Written test, oral test, oral presentation, group project, essay, etc.)

E. Learning Resources and Facilities

1. References and Learning Resources

Essential References	<p>دين، أو كلايفر (2002) الأمراض المنقولة بواسطة الغذاء ترجمة مسفر بن محمد الدقل و إسماعيل عيسى الشايب – مطابع جامعة الملك سعود – الرياض</p> <ul style="list-style-type: none"> هندي، مبد الحميد زيدان و إبراهيم، عبد المجيد محمد (1999) الملوثات البيئية و الكيمائية – الدار العربية للنشر- القاهرة المواصفات الدولية لنظام إدارة سلامة الغذاء بالمنشأة الغذائية ايزو 22000 – عام 2005 نشرات ومواصفات والخطوط الإرشادية الصادرة من هيئة دستور (CODEX) الأغذية
Supportive References	
Electronic Materials	<p>www.momra.gov.sa www.sFDA.gov.sa http://www.fsis.usda.gov/ http://www.food.gov.uk/safereating/ http://WWW.Codex.com</p>
Other Learning Materials	<ul style="list-style-type: none"> • Guide line and standards of codex • Standards of International Organization of standardization

2. Required Facilities and equipment

Items	Resources
facilities (Classrooms, laboratories, exhibition rooms, simulation rooms, etc.)	Lecture room equipped with blackboard, overhead projector, computer, and internet connection.

Items	Resources
Technology equipment (projector, smart board, software)	Data show, Smart Board, software, Internet connection
Other equipment (depending on the nature of the specialty)	Non

F. Assessment of Course Quality

Assessment Areas/Issues	Assessor	Assessment Methods
Effectiveness of teaching	Students	Indirect methods course evaluation survey
Effectiveness of students assessment	Faculty	Indirect
Quality of learning resources	Students	Indirect methods course evaluation survey
The extent to which CLOs have been achieved	Program Leaders	Direct through the tests
Other		

Assessor (Students, Faculty, Program Leaders, Peer Reviewer, Others (specify))

Assessment Methods (Direct, Indirect)

G. Specification Approval Data

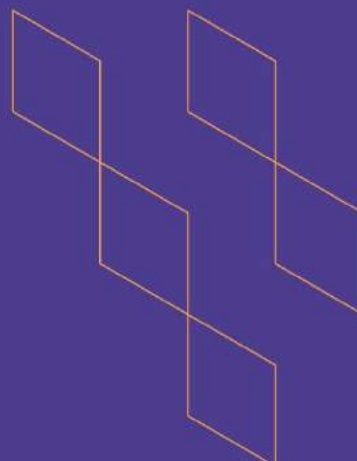
COUNCIL /COMMITTEE	COUNCIL OF VETERINARY MEDICINE DEPARTMENT
REFERENCE NO.	1444-10-95
DATE	19/2/2023





T-104
2022

Course Specification



Course Title: Surgical anatomy
Course Code: VMD 326
Program: Bachelor of Veterinary Medicine
Department: Veterinary Medicine
College: Agriculture and Veterinary Medicine
Institution: Qassim University
Version: T-104 (2022)
Last Revision Date: 1/2/2023



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A. General information about the course:

Course Identification	
1. Credit hours:	2 (1 + 1)
2. Course type	
a.	University <input type="checkbox"/> College <input type="checkbox"/> Department <input checked="" type="checkbox"/> Track <input type="checkbox"/> Others <input type="checkbox"/>
b.	Required <input checked="" type="checkbox"/> Elective <input type="checkbox"/>
3. Level/year at which this course is offered : Level 4 / Second year	
4. Course general Description The course includes studying the surgical anatomy of the various parts of the body in the animal, including knowledge and identification of nerve, block nerves in the head region , what is related to the eyes, ear, maxilla and mandible fractures, suturing wounds in the upper and lower lips area, and knowledge of the places that undergo most surgeries in the neck region, block nerves in the abdominal region, block nerves in the forelimb, block nerves sites in the hindlimb.	
5. Pre-requirements for this course (if any): 224 VMD	
6. Co- requirements for this course (if any): None	
7. Course Main Objective(s) <ul style="list-style-type: none"> • Provide the student with basic knowledge in locating nerves in various parts of the body to perform various surgical operations • The course prepares students for other medical sciences at other levels and other courses such as obstetric surgery 	

1. Teaching mode (mark all that apply)

No	Mode of Instruction	Contact Hours	Percentage
1.	Traditional classroom	60	100%
2.	E-learning		
3.	Hybrid <ul style="list-style-type: none"> • Traditional classroom • E-learning 		



No	Mode of Instruction	Contact Hours	Percentage
4.	Distance learning		

2. Contact Hours (based on the academic semester)

No	Activity	Contact Hours
1.	Lectures	15
2.	Laboratory/Studio	30
3.	Field	-
4.	Tutorial	-
5.	Others (specify)	-
	Total	45





B. Course Learning Outcomes (CLOs), Teaching Strategies and Assessment Methods

Code	Course Learning Outcomes	Code of CLOs aligned with program	Teaching Strategies	Assessment Methods
1.0	Knowledge and understanding			
1.1	Students will be able to plan for strategic methods used for disease prevention and control.	S3	- Lectures. - Practical sessions.	- One midterm exams. - Final practical exams - Final written exam.
1.2	Students will be able to interpret the available data in the field of disease prevention.	S5	- Using internet. - Individual presentation.	- Class activity discussion.
...				
2.0	Skills			
2.1				
2.2				
...				
3.0	Values, autonomy, and responsibility			
3.1	Students will be able to make independent and critical assessments	V3		- Class participation discussion.
3.2				
...				

C. Course Content

No	List of Topics	Contact Hours
1.	Introduction to the course	1
2.	Surgical anatomy of the head region	4
3.	Surgical anatomy of the neck region	2
4.	Surgical anatomy of the thoracic region	2
5.	Surgical anatomy of the abdomen region	2
6.	Surgical anatomy of the forelimb region	2
7.	Surgical anatomy of the hindlimb region	2





D. Students Assessment Activities

No	Assessment Activities *	Assessment timing (in week no)	Percentage of Total Assessment Score
1.	First and second midterm (written test).	7 & 12	20
2.	Class Participation discussion (Oral test).	Through semester	5
3.	Practical exam (written test).		25
4.	Final theoretical exams	15	50

*Assessment Activities (i.e., Written test, oral test, oral presentation, group project, essay, etc.)





E. Learning Resources and Facilities

1. References and Learning Resources

Essential References	Regional and Surgical Anatomy of Bovines, 2016
Supportive References	<ul style="list-style-type: none"> Journal of veterinary anatomy Anatomia, Histologia, Embryologia International Journal of Veterinary and Animal Veterinary Sciences
Electronic Materials	<ul style="list-style-type: none"> https://www.imaios.com/en/vet-anatomy
Other Learning Materials	

2. Required Facilities and equipment

Items	Resources
facilities (Classrooms, laboratories, exhibition rooms, simulation rooms, etc.)	<ul style="list-style-type: none"> Classroom of 20 students capacity. Laboratory of 15students capacity.
Technology equipment (projector, smart board, software)	Data show.
Other equipment (depending on the nature of the specialty)	Chemicals. Glasses and kits for experimental sessions.

F. Assessment of Course Quality

Assessment Areas/Issues	Assessor	Assessment Methods
Effectiveness of teaching	Students	Course Evaluation Survey (Indirect)
Effectiveness of students assessment	<ul style="list-style-type: none"> Students Program Leaders 	<ul style="list-style-type: none"> Course Evaluation Survey (Indirect). Result of the course (Direct)
Quality of learning resources	Students	Course Evaluation Survey (Indirect)





Assessment Areas/Issues	Assessor	Assessment Methods
The extent to which CLOs have been achieved	<ul style="list-style-type: none"> Students Instructor 	<ul style="list-style-type: none"> Course Evaluation Survey (Indirect). Checking students' performance in the test (Direct).
Other		

Assessor (Students, Faculty, Program Leaders, Peer Reviewer, Others (specify))

Assessment Methods (Direct, Indirect)

G. Specification Approval Data

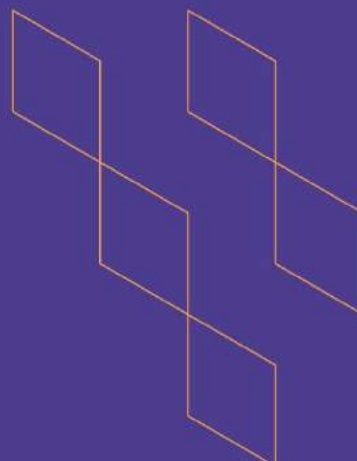
COUNCIL /COMMITTEE	COUNCIL OF VETERINARY MEDICINE DEPARTMENT
REFERENCE NO.	1444-10-95
DATE	19/2/2023





T-104
2022

Course Specification



Course Title: Nutrition and Immunity in Human
Course Code: FSNU 341
Program: Food Science & Human Nutrition program
Department: Food Science & Human Nutrition
College: College of Agric. & Veterinary Medicine
Institution: Qassim University
Version: T104 Course Specifications
Last Revision Date: V2022



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A. General information about the course:

Course Identification	
1. Credit hours:	2h (2+0)
2. Course type	
a.	University <input type="checkbox"/> College <input checked="" type="checkbox"/> Department <input type="checkbox"/> Track <input type="checkbox"/> Others <input type="checkbox"/>
b.	Required <input type="checkbox"/> Elective <input checked="" type="checkbox"/>
3. Level/year at which this course is offered:	Optional college-2022
4. Course general Description This course is intended to Qualify the student with Understand the relationship between nutrition and immune response, Know the effect of different nutrients on immune response, Know the effect of amino and fatty acids on immune response, understand how to reduce risks of disease through dietary modifications.	
5. Pre-requirements for this course (if any): BCH 301, Principal of Biochemistry	
6. Co- requirements for this course (if any): None	
7. Course Main Objective(s) <ul style="list-style-type: none"> • Qualify the students to Understand the relationship between nutrition and immune response. • Know the effect of different nutrients on immune response. • Summarize nutritional care and restricted diet for patients with Food sensitivities and auto immune diseases. 	

1. Teaching mode (mark all that apply)

No	Mode of Instruction	Contact Hours	Percentage
1.	Traditional classroom	30 hr.	100%
2.	E-learning		
3.	Hybrid <ul style="list-style-type: none"> • Traditional classroom • E-learning 		
4.	Distance learning		

2. Contact Hours (based on the academic semester)

No	Activity	Contact Hours
1.	Lectures	30 hr.
2.	Laboratory/Studio	
3.	Field	



4.	Tutorial	
5.	Others (specify)	
Total		30 hr.

B. Course Learning Outcomes (CLOs), Teaching Strategies and Assessment Methods

Code	Course Learning Outcomes	Code of CLOs aligned with program	Teaching Strategies	Assessment Methods
1.0	Knowledge and understanding			
1.1				
1.2	Introduces students to the most important principles governing the functions of the human immune system and to Understand the relationship between nutrition and immune response.	K2	Theoretical lectures Using pictures and power point Interactive learning process through questions and answers in class.	Pe, F,
...				
2.0	Skills			
2.1				
2.2	Summarize nutritional care and restricted diet for patients with Food sensitivities and auto immune diseases.	S2	Lectures are followed by numerous examples, some of which are practical in nature, to illustrate the application and use.	Pe, F, Hw,
...				
3.0	Values, autonomy, and responsibility			
3.1	Qualify the students to Understand the relationship between nutrition and immune response	V1	Theoretical lectures Using pictures and power point Interactive learning process through questions and answers in class.	Pe, F,
3.2				
...				



C. Course Content

No	List of Topics	Contact Hours
1.	Introduction to human immune system	2hr
2.	Nutritional status and immune response	2hr
3.	Deficiency of immune response among elderly and its relation with nutrition	2hr
4.	Nutritional factors that modify immune response	2hr
5.	Food allergy and intolerance	2hr
6.	Immunomodulatory Effect of macronutrients	2hr
7.	Immunomodulatory Effect of micronutrients	4hr
8.	Reduction of disease risks through dietary modifications	2hr
9.	The Immunomodulator roll of functional and nutraceuticals food	4hr
10.	Meal Planning for some different food allergy, intolerance and auto immune disease	8hr
Total		30hr.

D. Students Assessment Activities

No	Assessment Activities *	Assessment timing (in week no)	Percentage of Total Assessment Score
1.	Homework, activity and sharing	Along the semester	10%
2.	Periodical Exam	7-11 th week	30%
3.	Final Exam	16 -17 th week	60%
...			

*Assessment Activities (i.e., Written test, oral test, oral presentation, group project, essay, etc.)





E. Learning Resources and Facilities

1. References and Learning Resources

Essential References	Lillian, L. (1999): Nutrition and Immunity in Man. ILSI Europe, Belgium. Prakash Shetty (2010): Nutrition, Immunity and Infection. Cambridge University Press, Cambridge, UK.
Supportive References	Course materials
Electronic Materials	www.ift.org •WHO web site •AOAC on line
Other Learning Materials	None

2. Required Facilities and equipment

Items	Resources
facilities (Classrooms, laboratories, exhibition rooms, simulation rooms, etc.)	Lecture room contain at list 45 seats and equipped with a White board.
Technology equipment (projector, smart board, software)	Data show device, computer, or lab top and internet connection
Other equipment (depending on the nature of the specialty)	None

F. Assessment of Course Quality

Assessment Areas/Issues	Assessor	Assessment Methods
Effectiveness of teaching	Students	Indirect method
Effectiveness of students assessment	Faculty	Indirect method
Quality of learning resources	Students	Indirect method
The extent to which CLOs have been achieved	Program Leaders	Direct method
Other		

Assessor (Students, Faculty, Program Leaders, Peer Reviewer, Others (specify))

Assessment Methods (Direct, Indirect)

G. Specification Approval Data

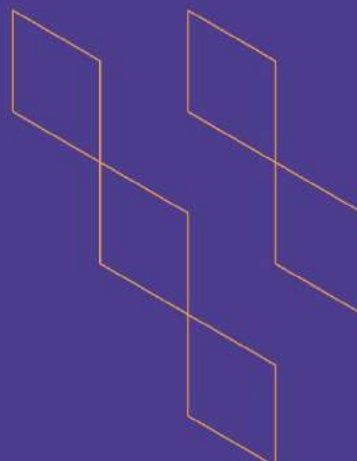
COUNCIL /COMMITTEE	COUNCIL OF VETERINARY MEDICINE DEPARTMENT
REFERENCE NO.	1444-10-95
DATE	19/2/2023





T-104
2022

Course Specification



Course Title: Molecular biology
Course Code: VMD 348
Program: Bachelor of Veterinary Medicine.
Department: Veterinary Medicine.
College: Agriculture and Veterinary Medicine.
Institution: Qassim University .
Version: T-104 (2022)
Last Revision Date: 1/2/2021 .





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F. Assessment of Course Quality	5
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A. General information about the course:

Course Identification	
1. Credit hours:	2 (1+ 1)
2. Course type	
a.	University <input type="checkbox"/> College <input checked="" type="checkbox"/> Department <input type="checkbox"/> Track <input type="checkbox"/> Others <input type="checkbox"/>
b.	Required <input type="checkbox"/> Elective <input checked="" type="checkbox"/>
3. Level/year at which this course is offered:	
4. Course general Description: This course is concerned with the study of components of the cell with more emphasis on the processes of DNA replication, transcription, translation, DNA damage and repairs, cell division as well as laboratory techniques utilized in molecular biology..	
5. Pre-requirements for this course (if any): Biochemistry (BCH 301).	
6. Co- requirements for this course (if any): NA	
7. Course Main Objective(s): 1. To provide students with an adequate knowledge on the basics of molecular biology & molecular tools used in diseases diagnosis.	

1. Teaching mode (mark all that apply)

No	Mode of Instruction	Contact Hours	Percentage
1.	Traditional classroom	45	100%
2.	E-learning		
3.	Hybrid <ul style="list-style-type: none"> • Traditional classroom • E-learning 		
4.	Distance learning		

2. Contact Hours (based on the academic semester)

No	Activity	Contact Hours
1.	Lectures	30
2.	Laboratory/Studio	15
3.	Field	-
4.	Tutorial	-
5.	Others (specify)	-
	Total	45





B. Course Learning Outcomes (CLOs), Teaching Strategies and Assessment Methods

Co de	Course Learning Outcomes	Code of CLOs aligned with program	Teaching Strategies	Assessment Methods
1.0	Knowledge and understanding			
1.1	Students will be able to list and recognize basic knowledge related to cell and molecular biology.	K1	Lectures contain tools currently used in molecular biology and their applications, pictures of Cell division and DNA replication, transcription and translation. Laboratory sessions about some molecular diagnostic methods. - Giving handouts. - Continuous revisions	1. Quarterly exam 2. Practical exam 3. Final exam
2.0	Skills			
2.1	Students will be familiar with widely used molecular diagnostic methods.	S1	Lectures contain tools currently used in molecular biology and their applications, pictures of Cell division and DNA replication, transcription and translation. Laboratory sessions about some molecular diagnostic methods. - Giving handouts. - Continuous revisions	1. Quarterly exam 2. Practical exam 3. Final exam
3.0	Values, autonomy, and responsibility			
3.1				

C. Course Content

No	List of Topics	Contact Hours
1.	General introduction to the cell	10
2.	Nucleic acids	5
3.	DNA replication	5
4.	Transcription and translation	5
5.	DNA damage and repair	5
6.	Cell division	5
7.	DNA isolation	5
8.	Tools currently used in molecular biology and their applications	5
Total		45





D. Students Assessment Activities

No	Assessment Activities *	Assessment timing (in week no)	Percentage of Total Assessment Score
1.	First quarterly exam	6	15
2.	Second quarterly exam	12	15
3.	Assignments	Anytime	5
4.	Practical exam	15	15
5.	Final exam	16	50

*Assessment Activities (i.e., Written test, oral test, oral presentation, group project, essay, etc.)

E. Learning Resources and Facilities

1. References and Learning Resources

Essential References	Molecular Biology of the Cell by Bruce Alberts
Supportive References	Journal of Veterinary Surgery
Electronic Materials	
Other Learning Materials	Suitable videos and animations from the internet.

2. Required Facilities and equipment

Items	Resources
facilities (Classrooms, laboratories, exhibition rooms, simulation rooms, etc.)	Air conditioned classroom of a minimum of 35 seats and powered by multimedia equipment.
Technology equipment (projector, smart board, software)	Data show connected to a computer (desktop or laptop), access to the world wide web.
Other equipment (depending on the nature of the specialty)	None

F. Assessment of Course Quality

Assessment Areas/Issues	Assessor	Assessment Methods
Effectiveness of teaching	Students – graduates- faculty – program administration – committee for quality assurance and accreditation	Students survey and questionnaire annual reports prepared by the program administrative committee
Effectiveness of students assessment	Students- Graduates- employers - committee for quality assurance and accreditation	Survey for students – graduates – employers – bi-annual reports of the quality assurance unit
Quality of learning resources	Students – graduates – peer reviewer	Surveys and periodical visits of internal and external per reviewers
The extent to which CLOs have been achieved	Students Instructor	Course Evaluation Survey (Indirect). Checking students'





Assessment Areas/Issues	Assessor	Assessment Methods
		performance in the test (Direct).
Other		

[Assessor](#) (Students, Faculty, Program Leaders, Peer Reviewer, Others (specify))

[Assessment Methods](#) (Direct, Indirect)

G. Specification Approval Data

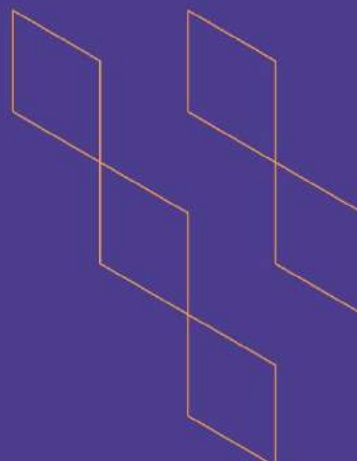
COUNCIL /COMMITTEE	VETERINARY DEPARTMENT
REFERENCE NO.	1444-10-95
DATE	19.02.2023





T-104
2022

Course Specification



Course Title: Tissue Culture
Course Code: VMD349
Program: Bachelor of Veterinary Medicine
Department: Department of Veterinary Medicine
College: College of Agriculture and Veterinary Medicine
Institution: Qassim University
Version: : T-104 (2022)
Last Revision Date: 1/2/2021



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A. General information about the course:

Course Identification	
1. Credit hours:	2 (1 + 1)
2. Course type	
a.	University <input type="checkbox"/> College <input type="checkbox"/> Department <input type="checkbox"/> Track <input type="checkbox"/> Others <input type="checkbox"/>
b.	Required <input type="checkbox"/> Elective <input checked="" type="checkbox"/>
3. Level/year at which this course is offered:	Level 3 and after
4. Course general Description Systemic histology is a discipline which provides students/beneficial with the structure and correlating functions of animal body systems using microscopy. The course involves the study of animal systems appearance and explores histologically and ultra-structurally the various organ systems including cardiovascular, lymphatic, integumentary (skin), digestive, respiratory, urinary, endocrine, male and female reproductive systems as well as special senses (eye and ear). Functional correlations will also be made.	
5. Pre-requirements for this course (if any): General histology (VMD222)	
6. Co- requirements for this course (if any): N/A	
7. Course Main Objective(s)	
<ul style="list-style-type: none"> To provide students with the basic knowledge about systemic histology: Different animal systems structures and functions as well as how they can be characterized. The course will prepare the students for histopathology in the coming levels. 	

1. Teaching mode (mark all that apply)

No	Mode of Instruction	Contact Hours	Percentage
1.	Traditional classroom	60	100%
2.	E-learning		
3.	Hybrid <ul style="list-style-type: none"> Traditional classroom E-learning 		
4.	Distance learning		

2. Contact Hours (based on the academic semester)

No	Activity	Contact Hours
1.	Lectures	30
2.	Laboratory/Studio	30
3.	Field	-
4.	Tutorial	-





5.	Others (specify)	-
	Total	60

B. Course Learning Outcomes (CLOs), Teaching Strategies and Assessment Methods

Code	Course Learning Outcomes	Code of CLOs aligned with program	Teaching Strategies	Assessment Methods
1.0	Knowledge and understanding			
1.1	Students will be able to list and recognize knowledge related to tissue culture	K1	- Lectures. - Practical sessions	-Two midterm exams. - Final written exam. - Practical exam. - Quizzes and classwork
1.2				
...				
2.0	Skills			
2.1				
2.2				
...				
3.0	Values, autonomy, and responsibility			
3.1				
3.2				

C. Course Content

No	List of Topics	Contact Hours
1.	Introduction	2
2.	Histology and cytology revision	2
3.	Sterilization techniques & types of animal tissue cultures	4
4.	Methods of preparation of tissue culture and requirements	7
5	Preparation of media and sera/ Culturing and sub-culturing of animal cells/ Cell counting and viability	3
6	Staining of animal cells/ Preservation of cells	6
7	Propagation and isolation of virus in tissue cells	3
8	Characterization of pathological lesions (cytopathic effects)	3
9	Propagation of viruses for vaccine production	9
10	Animal cell culture applications	6
	Total	60





D. Students Assessment Activities

No	Assessment Activities *	Assessment timing (in week no)	Percentage of Total Assessment Score
1.	First and second midterm (written test).	7 & 12	20
2.	Class work	Throughout semester	5
3.	Quiz	3	5
4.	Practical exam (written test).	15	20
5.	Final exam (written test)	16	50

*Assessment Activities (i.e., Written test, oral test, oral presentation, group project, essay, etc.)



E. Learning Resources and Facilities

1. References and Learning Resources

Essential References	- Culture of Animal Cells: A Manual of Basic Technique and Specialized Applications, 6th Edition. By: R. Ian Freshney, Wiley-Blackwell, 2010.
Supportive References	Print Journals and other books related to tissue cultures
Electronic Materials	Electronic Journals and books related to tissue cultures
Other Learning Materials	

2. Required Facilities and equipment

Items	Resources
facilities (Classrooms, laboratories, exhibition rooms, simulation rooms, etc.)	-Classroom of 20 student's capacity. -Laboratory of 15students capacity.
Technology equipment (projector, smart board, software)	-Projectors -Software provision on blackboard
Other equipment (depending on the nature of the specialty)	Chemicals. Glasses and kits for experimental sessions.

F. Assessment of Course Quality

Assessment Areas/Issues	Assessor	Assessment Methods
Effectiveness of teaching	Students	Course Evaluation Survey (Indirect)
Effectiveness of students assessment	Students Program Leaders	Course Evaluation Survey (Indirect). Result of the course (Direct)
Quality of learning resources	Students	Course Evaluation Survey (Indirect)
The extent to which CLOs have been achieved	Students Instructor	Course Evaluation Survey (Indirect). Exams (Direct).
Other		

Assessor (Students, Faculty, Program Leaders, Peer Reviewer, Others (specify))

Assessment Methods (Direct, Indirect)

G. Specification Approval Data

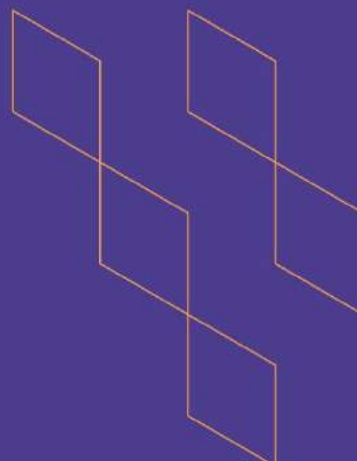
COUNCIL /COMMITTEE	COUNCIL OF VETERINARY MEDICINE DEPARTMENT
REFERENCE NO.	1444-10-95
DATE	19/2/2023





T-104
2022

Course Specification



Course Title: Functional Foods
Course Code: FSNU352
Program: Food Science and Human Nutrition
Department: Food Science and Human Nutrition
College: College of Agriculture and Veterinary Medicine
Institution: Qassim University
Version: Ver. 4
Last Revision Date: <i>Pick Revision Date.</i>



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E. Learning Resources and Facilities	6
1. References and Learning Resources	6
2. Required Facilities and Equipment	6
F. Assessment of Course Quality	6
G. Specification Approval Data	7



A. General information about the course:

Course Identification	
1. Credit hours:	2 h (1+1)
2. Course type	
a.	University <input type="checkbox"/> College <input checked="" type="checkbox"/> Department <input type="checkbox"/> Track <input type="checkbox"/> Others <input type="checkbox"/>
b.	Required <input type="checkbox"/> Elective <input checked="" type="checkbox"/>
3. Level/year at which this course is offered:	4 th , 5 th , and 6 th level
4. Course general Description This course will give students an introduction about functional foods (definition and concept development) and its positive impact on human health. Besides, the scientific basis, technologies of producing different types of functional foods. More focus will be on important examples of functional foods from dairy, cereals products. Uses of functional foods to reduce risk of chronic diseases (coronary heart diseases – diabetes – cancer – osteoporosis).	
5. Pre-requirements for this course (if any): FSNU344	
6. Co- requirements for this course (if any): Non	
7. Course Main Objective(s) The main objective of this course is to provide a comprehensive understanding of functional foods and their positive health effects. Students will learn about the different topics, such as phytochemicals, zoochemical (CLA, Omega3), probiotics, prebiotics, symbiotic, and their health benefits. In addition, the course will focus on the relationship between the bioactivity of key components in functional foods and their health benefits against some chronic diseases (CHD, Diabetes, osteoporosis, and cancers).	

1. Teaching mode (mark all that apply)

No	Mode of Instruction	Contact Hours	Percentage
1.	Traditional classroom	3	100
2.	E-learning		
3.	Hybrid <ul style="list-style-type: none"> • Traditional classroom • E-learning 		
4.	Distance learning		



2. Contact Hours (based on the academic semester)

No	Activity	Contact Hours
1.	Lectures	1h (1×15)
2.	Laboratory/Studio	2 h (2×15)
3.	Field	
4.	Tutorial	
5.	Others (specify)	
	Total	45

B. Course Learning Outcomes (CLOs), Teaching Strategies and Assessment Methods

Code	Course Learning Outcomes	Code of CLOs aligned with program	Teaching Strategies	Assessment Methods
1.0	Knowledge and understanding			
1.1	Explain functional food concepts, history and regulations.	K1	Lectures	F, Pe, Q
1.2	Describe the health benefits of different phytochemicals and their use against some chronic diseases.	K2	Lectures	F, Pe, Q
...				
2.0	Skills			
2.1	Determine different functional ingredients using qualitative and quantitative techniques.	S1	Laboratory	L, R
2.2				
...				
3.0	Values, autonomy, and responsibility			
3.1				
3.2				
...				

Pe: Periodical exams, F: final exam, R: Reports, Q: quiz, L: Practical Exam.

C. Course Content

No	List of Topics	Contact Hours
	A: Lectures	





1.	Introduction to functional foods, its categories and market.	1
2.	History and development of functional food concept.	2
3.	Regulation and health claims of functional foods.	2
4.	Phytochemicals in fruit and vegetables.	2
5.	Functional food products: Milk-based functional foods. Cereals-based functional foods.	3
6.	Functional foods for risk reduction of chronic diseases (coronary heart diseases – diabetes – cancers - osteoporosis).	5
<u>B: Practical</u>		
7.	Extraction of phytochemicals from plant sources	6
8.	Sprouting and germination of seeds and its health potentials	2
9.	Detection of phytochemicals – Qualitative analysis	4
10.	Detection of phytochemicals – Quantitative analysis	4
11.	Determination of total phenolic acids.	2
12.	Determination of total flavonoids.	2
13.	Detailed studies on probiotic, prebiotic and symbiotic.	2
14.	Determination of antioxidant activity – DPPH method	2
15.	Determination of antioxidant activity – ABTS method	2
16.	Determination of Chlorophyll and beta carotene .	2
17.	Determination of vitamin C.	2
Total		45

D. Students Assessment Activities

No	Assessment Activities *	Assessment timing (in week no)	Percentage of Total Assessment Score
1.	Periodical exams	6-7	20
2.	Practical Exam	15	20
3.	Quiz	In some weeks	5
4.	Reports	In some weeks	5
4.	Final exam	16-17	50

*Assessment Activities (i.e., Written test, oral test, oral presentation, group project, essay, etc.)





E. Learning Resources and Facilities

1. References and Learning Resources

Essential References	<ul style="list-style-type: none"> Smith, J. Q and Charter, E. (2010). "Functional food product development". Blackwell publishing Ltd., United Kingdom (Available in the central library of QU). Schmidl, M. K. and Labuza, T. P. (2000). Essentials of functional foods. Aspen Publication, Inc., Gaithersburg, Maryland (Available in the central library of QU).
Supportive References	<ul style="list-style-type: none"> Margaret Ashwell, (2002): Concepts of Functional Foods. ILSI Europe, Brussels, Belgium Gibson, G. R., and Williams, C. R. (2000): Functional foods: concept to product. CRC Press LLC, Boca Raton. USA. Chadwick, R., Henson, S., Moseley, B., Koenen, G., Liakopoulos, M., Midden, C., Palou, A., Rechkemmer, G., Schröder, D., Wright, A. von. (2003). "Functional Foods". Springer-Verlag, Berlin, Germany.
Electronic Materials	http://www.journals.elsevier.com/journal-of-functional-foods/
Other Learning Materials	Course materials provided to students at the beginning of the semester, which containing all PowerPoint slides of all topics.

2. Required Facilities and equipment

Items	Resources
facilities (Classrooms, laboratories, exhibition rooms, simulation rooms, etc.)	Classrooms, laboratory
Technology equipment (projector, smart board, software)	Projector and computer
Other equipment (depending on the nature of the specialty)	Rotary evaporators, chemicals and reagents required for practical work.

F. Assessment of Course Quality

Assessment Areas/Issues	Assessor	Assessment Methods
Effectiveness of teaching	Students	Indirect method: Course Evaluation Survey
Effectiveness of students assessment	Students	Indirect method: Course Evaluation Survey
Quality of learning resources	Students	Indirect method: Course Evaluation Survey
The extent to which CLOs have been achieved	Course instructors	Direct method: Students achievements in





Assessment Areas/Issues	Assessor	Assessment Methods
		assessment activities
Other		

Assessor (Students, Faculty, Program Leaders, Peer Reviewer, Others (specify))

Assessment Methods (Direct, Indirect)

G. Specification Approval Data

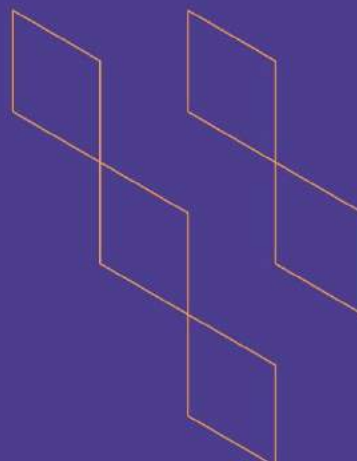
COUNCIL /COMMITTEE	COUNCIL OF VETERINARY MEDICINE DEPARTMENT
REFERENCE NO.	1444-10-95
DATE	19/2/2023





T-104

Course Specification



Course Title: Production of ornamental animals and birds
Course Code: APP 380
Program: Animal production
Department: Animal production and breeding
College: Agriculture and veterinary medicine
Institution: Qassim university
Version: Course Specification Version Number
Last Revision Date: Feb 2023



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A. General information about the course:

Course Identification

1. Credit hours:

2. Course type

a. University College Department Track Others

b. Required Elective

3. Level/year at which this course is offered: 3

4. Course general Description

This course deals with the care, feeding, reproduction and inheritance of ornamental animals and birds

5. Pre-requirements for this course (if any): APP 211

6. Co- requirements for this course (if any):

7. Course Main Objective(s)

Teaching the students the Breeds, housing, nutrition, reproduction, management and rearing of ornamental animals

1. Teaching mode (mark all that apply)

No	Mode of Instruction	Contact Hours	Percentage
1.	Traditional classroom	45	100
2.	E-learning		
3.	Hybrid <ul style="list-style-type: none"> • Traditional classroom • E-learning 		
4.	Distance learning		





2. Contact Hours (based on the academic semester)

No	Activity	Contact Hours
1.	Lectures	30
2.	Laboratory/Studio	
3.	Field	
4.	Tutorial	
5.	Others (specify)	15
	Total	45



B. Course Learning Outcomes (CLOs), Teaching Strategies and Assessment Methods

Code	Course Learning Outcomes	Code of CLOs aligned with program	Teaching Strategies	Assessment Methods
1.0	Knowledge and understanding			
1.1	Identify the breeds of ornamental animals and birds		Lectures and practical lessons	periodical exam
1.2	Learn about the feeding and reproduction of ornamental animals and birds		Lectures and practical lessons	periodical exam
...				
2.0	Skills			
2.1	Learn in animal care and nutrition		Practical lessons	Periodic and final exam
2.2				
...				
3.0	Values, autonomy, and responsibility			
3.1	Graduate familiar with animal ethics and welfare		Lectures	Periodic and final exams
3.2	A graduate interested in preserving endangered animals		Lectures	Periodic and final exams
...				

C. Course Content

No	List of Topics	Contact Hours
1.	Breeds of ornamental animals (cats, dogs, rabbits, fish, hamsters, turtles) - Breeds of ornamental birds (canaries, parrots, pigeons, pigeons, ducks)	8
2.	Shelter systems for ornamental animals and birds	4
3.	Feeding ornamental animals and birds	8
4.	Breeding of ornamental animals and birds	4
5.	Breeding and management of ornamental animals and birds	4



6.	Environmental conditions, ornamental animals and birds	5
7.	Marketing of ornamental animals and birds	4
8.	Knowledge of the structural and administrative structure of zoos	8
Total		45

D. Students Assessment Activities

No	Assessment Activities *	Assessment timing (in week no)	Percentage of Total Assessment Score
1.	Periodical and practical tests	5, 10, 15	35%
2.	Participation in the lecture halls	8,10,12, 14	5%
3.	Quizzes	4	5%
4.	Field reports	15	5%
5.	Final Exams	16	50%
6.	Periodical and practical tests	5, 10, 15	35%

*Assessment Activities (i.e., Written test, oral test, oral presentation, group project, essay, etc.)



E. Learning Resources and Facilities

1. References and Learning Resources

Essential References	Encyclopedia of Aquarium & Pond Fish , Alderton 2008.
Supportive References	<p>كتاب "من عجائب الخلق في عالم الطيور" ، محمد اسماعيل جاويش (الدار الذهبية كتاب "كلب بلدي قط شيرازي" عن الحيوانات الأليفة في مصر، أحمد النبراوي (دار (المصري للنشر والتوزيع).</p> <p>Books:</p> <p>1-Caring for Your Canary. Paul Ruddock , Paradise Press.</p> <p>2- The Alpaca Breeding Book. K. D. Galbraith, Walnut Creek Publishing 2013.</p>
Electronic Materials	<p>Waterfowls Management Guidelines (wildpro.twycrosszoo.org)</p> <ul style="list-style-type: none"> • Poultry web site
Other Learning Materials	N/A

2. Required Facilities and equipment

Items	Resources
<p>facilities</p> <p>(Classrooms, laboratories, exhibition rooms, simulation rooms, etc.)</p>	N/A
<p>Technology equipment</p> <p>(projector, smart board, software)</p>	
<p>Other equipment</p> <p>(depending on the nature of the specialty)</p>	

F. Assessment of Course Quality

Assessment Areas/Issues	Assessor	Assessment Methods
Effectiveness of teaching		
Effectiveness of students assessment		
Quality of learning resources		
The extent to which CLOs have been achieved		




Assessment Areas/Issues	Assessor	Assessment Methods
Other		

Assessor (Students, Faculty, Program Leaders, Peer Reviewer, Others (specify))

Assessment Methods (Direct, Indirect)

G. Specification Approval Data

COUNCIL /COMMITTEE	The report has been revised and approved by the departmental council.	
REFERENCE NO.		
DATE	20/3/2023	





T-104
2022

Course Specification

Course Title: Technical Management of Animal Production Farms
Course Code: APP 381
Program: Animal Production And Breeding Program/ Animal production TracTechk
Department: Animal Production And Breeding
College: Agriculture and Veterinary Medicine
Institution: Qassim University
Version: 3 rd Version
Last Revision Date: Feb 3, 2023



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1. References and Learning Resources	7
2. Required Facilities and Equipment	7
F. Assessment of Course Quality	7
G. Specification Approval Data	7

A. General information about the course:

Course Identification				
1. Credit hours:				
2. Course type				
a.	University <input type="checkbox"/>	College X	Department <input type="checkbox"/>	Track <input type="checkbox"/> Others <input type="checkbox"/>
b.	Required <input type="checkbox"/>	Elective X		
3. Level/year at which this course is offered: All Levels for all Departments.				
4. Course general Description				
5. Pre-requirements for this course (if any):				
6. Co- requirements for this course (if any): None				
7. Course Main Objective(s)				
The ability to provide students with an appropriate understanding of the technical management of animal production farms.				
At the end of the course, the student will be able to technically manage animal production farms.				

1. Teaching mode (mark all that apply)

No	Mode of Instruction	Contact Hours	Percentage
1.	Traditional classroom	45	100%
2.	E-learning	---	---
3.	Hybrid <ul style="list-style-type: none"> • Traditional classroom • E-learning 	---	----
4.	Distance learning	---	----

2. Contact Hours (based on the academic semester)

No	Activity	Contact Hours
1.	Lectures	15
2.	Laboratory/Studio	30
3.	Field	---
4.	Tutorial	---
5.	Others (specify)	
Total		45

B. Course Learning Outcomes (CLOs), Teaching Strategies and Assessment Methods

Code	Course Learning Outcomes	Code of CLOs aligned with program	Teaching Strategies	Assessment Methods
1.0	Knowledge and understanding			
1.1	The ability to provide students with an appropriate understanding of the	K1	1. Lecture 2. Role Playing	1,2,3, and 4

Code	Course Learning Outcomes	Code of CLOs aligned with program	Teaching Strategies	Assessment Methods
	technical management of animal production farms.			
1.2				
...				
2.0	Skills			
2.1	<p>At the end of the course the student should be able to:</p> <p>At the end of the course, the student will be able to technically manage animal production farms.</p>	S5	<p>1. Lecture</p> <p>2. Debate</p> <p>3. Brainstorming</p>	1,2,3, and4
2.2	<p>At the end of the course the student should be able to:</p> <p>select the type of activity and the decision regarding the size and the quality of activity in terms of the number and required investments, as well as the breeding and management system for the project that is chosen and the importance of biosecurity in such projects.</p>	S6	<p>1. Lecture</p> <p>2. Debate</p> <p>3. Brainstorming</p>	1,2,3, and 4
...				
3.0	Values, autonomy, and responsibility			
3.1				
3.2				
...				

Program Learning Outcomes

Program Learning Outcomes	
	<i>At the end of the program, the student should be able to:</i>
K1	Knowing the elements of production on the farm, types and functions of management.
K2	Understanding the fundamentals of management in animal production, recipes and characteristics of a successful farm manager.
K3	Recognize the types and classifications of decisions facing the farm manager.
K4	Describe the farm production costs.

K5	Describes Some of the main productive activities in animal production projects.
K6	
S1	Evaluate the role of animal health and biosecurity procedures.
S2	Represent the importance of farm records.
S3	Explain the role of motivation and incentives in farm management.
S4	The ability to provide students with an appropriate understanding of the technical management of animal production farms.
S5	At the end of the course, the student will be able to technically manage animal production farms.
S6	
S7	
V1	Illustrate the ability to work in teams, communicate with others, and take responsibility.
V2	Write reports that cover different aspects of technical management of animal production farms. in a correct way linguistically and scientifically

List of assessment methods

Assessment Methods			
No.	Tool	No.	Tool
1	Mid-term exam	6	Homework
2	Practical Exam	7	Presentations
3	Final exam	8	Reports
4	Participation in lecture rooms	9	Practical experiments and training
5	Computer skills		

C. Course Content

No	List of Topics	Contact Hours
1.		
2.		

Total		

D. Students Assessment Activities

No	Assessment Activities *	Assessment timing (in week no)	Percentage of Total Assessment Score
1.	First Mid-term exam	7 th week	10%
2.	Second Mid-term exam	10 th week	10%

No	Assessment Activities *	Assessment timing (in week no)	Percentage of Total Assessment Score
3.	Participation in lecture rooms	Every week	Bonus with a maximum value of 5 degree
4.	Practical Exam	8 th week	12.5%
5.	Report	13 th week	12.5%
6.	Final Exam	15 th week	50%

*Assessment Activities (i.e., Written test, oral test, oral presentation, group project, essay, etc.)

E. Learning Resources and Facilities

1. References and Learning Resources

Essential References	م.2016د. إيهاب الضمان، إدارة مزارع الإنتاج الحيواني، جامعة حماة ، سوريا ،
Supportive References	وزارة البيئة والمياه والزراعة، دليل الأمن الحيوي في المنشآت الزراعية، مركز وقاء، الرياض
Electronic Materials	
Other Learning Materials	

2. Required Facilities and equipmen◆

Items	Resources
facilities (Classrooms, laboratories, exhibition rooms, simulation rooms, etc.)	Classrooms for about 40 students
Technology equipment (projector, smart board, software)	
Other equipment (depending on the nature of the specialty)	

F. Assessment of Course Quality

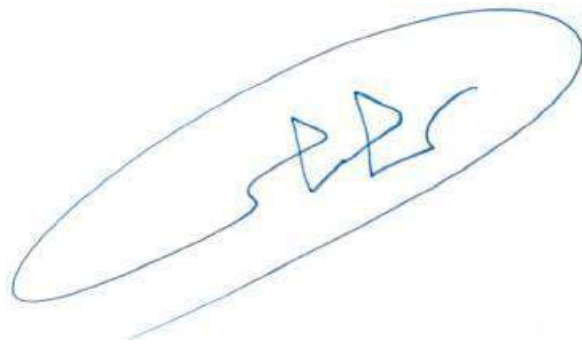
Assessment Areas/Issues	Assessor	Assessment Methods
Effectiveness of teaching	Students	Indirect Assessment (Questionnaire)
Effectiveness of students assessment	Students	Indirect Assessment (Questionnaire)
Quality of learning resources	Students	Indirect Assessment (Questionnaire)
The extent to which CLOs have been achieved	Students	Indirect Assessment (Questionnaire)
Other	-----	-----

Assessor (Students, Faculty, Program Leaders, Peer Reviewer, Others (specify))

Assessment Methods (Direct, Indirect)

G. Specification Approval Data

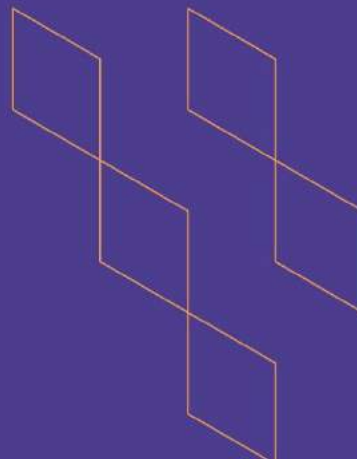
COUNCIL /COMMITTEE	The report has been revised and approved by the departmental council.
REFERENCE NO.	
DATE	20/3/2023





T-104
2022

Course Specification



Course Title: Veterinary Scientific Terms .
Course Code: BVM 413.
Program: Bachelor of Veterinary Medicine.
Department: Veterinary Medicine.
College: Agriculture and Veterinary Medicine.
Institution: Qassim University .
Version: T-104 (2022)
Last Revision Date: 1/2/2021 .





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2. Required Facilities and Equipment	4
F. Assessment of Course Quality	5
G. Specification Approval Data	5



A. General information about the course:

Course Identification	
1. Credit hours:	(1)
2. Course type	
a.	University <input type="checkbox"/> College <input type="checkbox"/> Department <input checked="" type="checkbox"/> Track <input type="checkbox"/> Others <input type="checkbox"/>
b.	Required <input checked="" type="checkbox"/> Elective <input type="checkbox"/>
3. Level/year at which this course is offered: Level 9/ Fifth year	
4. Course general Description: This course is concerned with the study of Veterinary Medical Terms.	
5. Pre-requirements for this course (if any): NA	
6. Co- requirements for this course (if any): NA	
7. Course Main Objective(s): Teaching the students veterinary medical terms.	

1. Teaching mode (mark all that apply)

No	Mode of Instruction	Contact Hours	Percentage
1.	Traditional classroom	12	100%
2.	E-learning		
3.	Hybrid <ul style="list-style-type: none"> • Traditional classroom • E-learning 		
4.	Distance learning		

2. Contact Hours (based on the academic semester)

No	Activity	Contact Hours
1.	Lectures	12
2.	Laboratory/Studio	-
3.	Field	-
4.	Tutorial	-
5.	Others (specify)	-
	Total	12

B. Course Learning Outcomes (CLOs), Teaching Strategies and Assessment Methods

Co de	Course Learning Outcomes	Code of CLOs aligned with program	Teaching Strategies	Assessment Methods
-------	--------------------------	-----------------------------------	---------------------	--------------------





Co de	Course Learning Outcomes	Code of CLOs aligned with program	Teaching Strategies	Assessment Methods
1.0	Knowledge and understanding			
1.1	Student will be able to list and recognize knowledge related to veterinary scientific terms	K1	Lectures contain medical terms, pictures of clinical cases, and examples of diseases.	1. quarterly exam 2. final exam
2.0	Skills			
2.5	Students will be able to demonstrate critical analysis of new information and research findings relevant to veterinary medicine.	S5	Lectures contain medical terms, pictures of clinical cases, and examples of diseases.	1. quarterly exam 2. final exam
3.0	Values, autonomy, and responsibility			
3.1				

C. Course Content

No	List of Topics	Contact Hours
1.	General scientific terms.	2
2.	Scientific terms of the digestive system	2
3.	Scientific terms of the respiratory system	2
4.	Scientific terms of the cardiovascular system	2
5.	Scientific terms of the urogenital system	2
6.	Scientific terms of the nervous system/ Ophthalmic scientific terms	2
Total		12

D. Students Assessment Activities

No	Assessment Activities *	Assessment timing (in week no)	Percentage of Total Assessment Score
1.	First quarterly exam	5	25
2.	Second quarterly exam	10	25
4	Final exam	16	50

*Assessment Activities (i.e., Written test, oral test, oral presentation, group project, essay, etc.)

E. Learning Resources and Facilities

1. References and Learning Resources

Essential References	
Supportive References	
Electronic Materials	Power point lectures
Other Learning Materials	prepared note

2. Required Facilities and equipment

Items	Resources
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Items	Resources
facilities (Classrooms, laboratories, exhibition rooms, simulation rooms, etc.)	Air conditioned classroom of a minimum of 35 seats and powered by multimedia equipment.
Technology equipment (projector, smart board, software)	Data show connected to a computer (desktop or laptop), access to the world wide web.
Other equipment (depending on the nature of the specialty)	

F. Assessment of Course Quality

Assessment Areas/Issues	Assessor	Assessment Methods
Effectiveness of teaching	Students – graduates- faculty – program administration – committee for quality assurance and accreditation	Students survey and questionnaire annual reports prepared by the program administrative committee
Effectiveness of students assessment	Students- Graduates- employers - committee for quality assurance and accreditation	Survey for students – graduates – employers – bi-annual reports of the quality assurance unit
Quality of learning resources	Students – graduates – peer reviewer	Surveys and periodical visits of internal and external per reviewers
The extent to which CLOs have been achieved	Students Instructor	Course Evaluation Survey (Indirect). Checking students' performance in the test (Direct).
Other		

Assessor (Students, Faculty, Program Leaders, Peer Reviewer, Others (specify))

Assessment Methods (Direct, Indirect)

G. Specification Approval Data

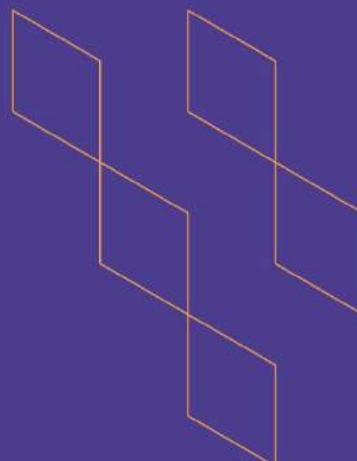
COUNCIL /COMMITTEE	VETERINARY DEPARTMENT
REFERENCE NO.	
DATE	





T-104
2022

Course Specification



Course Title: Clinical Microbiology
Course Code: VMD 456
Program: Bachelor of Veterinary Medicine
Department: Veterinary Medicine
College: Agriculture and Veterinary Medicine
Institution: Qassim University
Version: T-104 (2022)
Last Revision Date: 10/6/2022



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1. References and Learning Resources	6
2. Required Facilities and Equipment	6
F. Assessment of Course Quality	6
G. Specification Approval Data	7



A. General information about the course:

Course Identification	
1. Credit hours:	2 (1+1)
2. Course type	
a.	University <input type="checkbox"/> College <input type="checkbox"/> Department <input checked="" type="checkbox"/> Track <input type="checkbox"/> Others <input type="checkbox"/>
b.	Required <input type="checkbox"/> Elective <input checked="" type="checkbox"/>
3. Level/year at which this course is offered: Levels 7-10/4 th & 5 th year	
4. Course general Description:	
Diagnostic microbiology laboratory requirements, collection and submission of appropriate diagnostic samples for various pathological conditions and diseases. Microbial pathogens: microscopy, culture, serology and molecular characteristics.	
5. Pre-requirements for this course (if any):	
VMD 342 and VMD 344 (Introductory Microbiology Course and Virology Course)	
6. Co- requirements for this course (if any):	
None	
7. Course Main Objective(s)	
<ul style="list-style-type: none"> • Students will have enough knowledge about collection and submission of diagnostic specimens for identification of microbes. • Students will acquire the ability to identify microbes in clinical samples according to their microscopic, cultural, biochemical, serological and molecular characteristics. 	

1. Teaching mode (mark all that apply)

No	Mode of Instruction	Contact Hours	Percentage
1.	Traditional classroom	45	100
2.	E-learning		
3.	Hybrid <ul style="list-style-type: none"> • Traditional classroom • E-learning 		
4.	Distance learning		



2. Contact Hours (based on the academic semester)

No	Activity	Contact Hours
1.	Lectures	15
2.	Laboratory/Studio	30
3.	Field	
4.	Tutorial	
5.	Others (specify)	
Total		45

B. Course Learning Outcomes (CLOs), Teaching Strategies and Assessment Methods

Code	Course Learning Outcomes	Code of CLOs aligned with program	Teaching Strategies	Assessment Methods
1.0	Knowledge and understanding			
1.1	Students will be able to cope with recent knowledge of collection and submission of diagnostic specimens.	K2	<ul style="list-style-type: none"> - Lectures. - Assignments 	<ul style="list-style-type: none"> - Quizzes. - 1st & 2nd mid-term tests. - Assignments. - Final theory exam.
1.2	Students will be able to understand advances in diagnostic methods of microbial infections.			
2.0	Skills			
2.1	Students will be able to apply knowledge and biosafety measures pertaining handling and processing of clinical samples.	S1	Practical sessions: students will have clinical samples and know how to properly handle and process them for identification of microbes.	<ul style="list-style-type: none"> - Practical quizzes. - Final practical exam. - 1st & 2nd mid-term tests and final theory exam will contain some questions to assess this part.
2.2	Students will be able to determine and perform appropriate methods for identification of microbes in clinical samples.			
3.0	Values, autonomy, and responsibility			





Code	Course Learning Outcomes	Code of CLOs aligned with program	Teaching Strategies	Assessment Methods
3.1				
3.2				

C. Course Content

No	List of Topics	Contact Hours
1	Collection of various types of diagnostic samples.	4
2	Sample submission and submission forms.	4
3	Microscopic identification of microbes.	4
4	Cultural characteristics of microbes.	8
5	Biochemical identification of bacteria and fungi.	8
6	Serological identification of microbes.	5
7	Molecular characterization of microbes.	8
8	Interpretation of laboratory results of microbial identification.	4
Total		45

D. Students Assessment Activities

No	Assessment Activities *	Assessment timing (in week no)	Percentage of Total Assessment Score
1.	Quiz 1	3	2
2.	First mid-term test	6	10
3.	Quiz 2	8	2
4.	Practical test	9	3
5.	Second mid-term test	12	10
6.	Assignment and presentation	Any time	3
7.	Practical exam	14	20
8.	Final exam	16	50

*Assessment Activities (i.e., Written test, oral test, oral presentation, group project, essay, etc.)

E. Learning Resources and Facilities

1. References and Learning Resources

Essential References

- Patricia M. Tille (2021). Bailey & Scott's Diagnostic Microbiology, 15th Edition. Elsevier publisher.



	<ul style="list-style-type: none"> • Connie R. Mahon and Donald C. Lehman (2018). Textbook of Diagnostic Microbiology. Saunders Publisher. • Bryan Markey, Finola Leonard, Marie Archambault, Ann Cullinane, and Dores Maguire (2013). Clinical Veterinary Microbiology, 2nd ed, Mosby, Elsevier.
Supportive References	<ul style="list-style-type: none"> • Journals of Microbiology.
Electronic Materials	<ul style="list-style-type: none"> • Veterinary and public health web sites: FAO, OIE, CDC, WHO,...etc.
Other Learning Materials	

2. Required Facilities and equipment

Items	Resources
facilities (Classrooms, laboratories, exhibition rooms, simulation rooms, etc.)	<ul style="list-style-type: none"> • A well-furnished lecture room with a capacity of 30 students. • A laboratory with a capacity of 30 students.
Technology equipment (projector, smart board, software)	<ul style="list-style-type: none"> • Data show (projector and screen).
Other equipment (depending on the nature of the specialty)	<ul style="list-style-type: none"> • The laboratory should contain basic equipment (glass- and plastic-ware, autoclave, oven, incubator, microscopes).

F. Assessment of Course Quality

Assessment Areas/Issues	Assessor	Assessment Methods
Effectiveness of teaching	<ul style="list-style-type: none"> • Students • Program chairman 	<ul style="list-style-type: none"> • Course Evaluation Survey. • Field Experience Survey. • Program Evaluation Survey. • Faculty Evaluation Form (Self-assessment and assessment by the head of department) • Frequency of grades distribution form.
Effectiveness of students assessment	<ul style="list-style-type: none"> • Program Committee of Development and Quality 	<ul style="list-style-type: none"> • Internal moderation (form of frequency of grades distribution, unusual results)



Assessment Areas/Issues	Assessor	Assessment Methods
	<ul style="list-style-type: none"> • Students • Program chairman 	report, criteria used to revise question papers and random samples of student answered sheets). <ul style="list-style-type: none"> • Course Evaluation Survey. • Benchmarking with similar other course(s).
Quality of learning resources	<ul style="list-style-type: none"> • Students 	<ul style="list-style-type: none"> • Course Evaluation Survey.
The extent to which CLOs have been achieved	<ul style="list-style-type: none"> • Students • Instructor 	<ul style="list-style-type: none"> • Course Evaluation Survey. • Students' performance in the tests and tasks. • Field Experience Survey.
Other		

Assessor (Students, Faculty, Program Leaders, Peer Reviewer, Others (specify))

Assessment Methods (Direct, Indirect)

G. Specification Approval Data

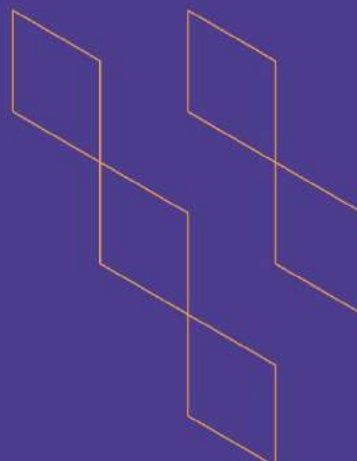
COUNCIL /COMMITTEE	COUNCIL OF VETERINARY MEDICINE DEPARTMENT
REFERENCE NO.	1444-10-95
DATE	19/2/2023





T-104
2022

Course Specification



Course Title: Veterinary Vaccines
Course Code: VMD 457.
Program: Bachelor of Veterinary Medicine.
Department: Veterinary Medicine.
College: Agriculture and Veterinary Medicine.
Institution: Qassim University .
Version: T-104 (2022)
Last Revision Date: 1/2/2021 .



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F. Assessment of Course Quality	5
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A. General information about the course:

Course Identification	
1. Credit hours:	2 (1+ 1)
2. Course type	
a.	University <input type="checkbox"/> College <input type="checkbox"/> Department <input checked="" type="checkbox"/> Track <input type="checkbox"/> Others <input type="checkbox"/>
b.	Required <input type="checkbox"/> Elective <input checked="" type="checkbox"/>
3. Level/year at which this course is offered:	
4. Course general Description: This course is concerned with the study of history and evolution of vaccinology, Vaccine types and characteristics, and Vaccination programs & evaluation.	
5. Pre-requirements for this course (if any): 343 VMD, 344 VMD.	
6. Co- requirements for this course (if any): NA	
7. Course Main Objective(s):	
1. Providing students with the basic knowledge of vaccines and immune sera (their evolution, types, production methods, preservation and their role in stimulating the immune system and preventing diseases).	
2. Enabling students to acquire the knowledge and practice of evaluation of vaccines and vaccination programs.	

1. Teaching mode (mark all that apply)

No	Mode of Instruction	Contact Hours	Percentage
1.	Traditional classroom	45	100%
2.	E-learning		
3.	Hybrid <ul style="list-style-type: none"> • Traditional classroom • E-learning 		
4.	Distance learning		

2. Contact Hours (based on the academic semester)

No	Activity	Contact Hours
1.	Lectures	30
2.	Laboratory/Studio	15
3.	Field	-
4.	Tutorial	-
5.	Others (specify)	-
	Total	45





B. Course Learning Outcomes (CLOs), Teaching Strategies and Assessment Methods

Co de	Course Learning Outcomes	Code of CLOs aligned with program	Teaching Strategies	Assessment Methods
1.0	Knowledge and understanding			
1.1	Students will be able to list and recognize basic knowledge related to veterinary vaccines.	K1	Lectures, accompanied with illustrating aids such as videos and animations. Giving handouts. Continuous revisions	1. Quarterly exam 2. Practical exam 3. Final exam
2.0	Skills			
2.1	Students will be able to recognize and perform laboratory methods to produce experimental vaccines and evaluate them	S1	- Laboratory preparation of experimental vaccines. - Giving the students some vaccines to evaluate. - Assignment discussions.	1. Quarterly exam 2. Practical exam 3. Final exam
3.0	Values, autonomy, and responsibility			
...				

C. Course Content

No	List of Topics	Contact Hours
1.	History and evolution of vaccinology.	10
2.	Vaccine types and characteristics	5
3.	Adjuvants (types & mechanism of action)	5
4.	Vaccines manufacture and modern techniques	5
5.	Vaccine preservation, preparation for injection & routes of injection.	5
6.	Vaccination programs & evaluation	5
7.	Evaluation of experimental and commercial vaccines	5
8.	Immunogenicity of vaccines	5
9.	Evaluation of immune response	5
Total		45

D. Students Assessment Activities

No	Assessment Activities *	Assessment timing (in week no)	Percentage of Total Assessment Score
1.	First quarterly exam	6	15
2.	Second quarterly exam	12	15
3.	Assignment & presentation	Any time	5
4.	Practical exam	15	15





No	Assessment Activities *	Assessment timing (in week no)	Percentage of Total Assessment Score
4	Final exam	16	50

*Assessment Activities (i.e., Written test, oral test, oral presentation, group project, essay, etc.)

E. Learning Resources and Facilities

1. References and Learning Resources

Essential References	Vaccinology – an Essential Guide. By: Gregg N. Milligan and Alan D.T. Barrett. John Wiley & Sons Ltd. - 2014. Do Vaccines Cause That?! A Guide for Evaluating Vaccine Safety Concerns. Martin G. Myers MD (Author), Diego Pineda. Paperback – 2008. Handouts are prepared especially for this course, translated from the appropriate textbooks in the field. These handouts are available at the Faculty photocopying center.
Supportive References	
Electronic Materials	Suitable videos and animations from the websites.
Other Learning Materials	computer-based programs/CD, professional standards or regulations and software. NA

2. Required Facilities and equipment

Items	Resources
facilities (Classrooms, laboratories, exhibition rooms, simulation rooms, etc.)	Air conditioned classroom of a minimum of 35 seats and powered by multimedia equipment. Equipped Veterinary clinic or Veterinary Teaching Hospital.
Technology equipment (projector, smart board, software)	Data show connected to a computer (desktop or laptop), access to the world wide web.
Other equipment (depending on the nature of the specialty)	Equipment for sterilization, incubation and refrigeration (autoclave, incubators, hot-air oven, refrigerator/freezer), ELISA reader, laminar flow hood, sensitive balance, hot plate/stirrer, glassware.

F. Assessment of Course Quality

Assessment Areas/Issues	Assessor	Assessment Methods
Effectiveness of teaching	Students – graduates- faculty – program administration – committee for quality assurance and accreditation	Students survey and questionnaire annual reports prepared by the program administrative committee
Effectiveness of students assessment	Students- Graduates- employers - committee for quality assurance and accreditation	Survey for students – graduates – employers – bi-annual reports of the quality assurance unit
Quality of learning resources	Students – graduates – peer reviewer	Surveys and periodical visits of internal and external per reviewers
The extent to which CLOs have	Students	Course Evaluation Survey





Assessment Areas/Issues	Assessor	Assessment Methods
been achieved	Instructor	(Indirect). Checking students' performance in the test (Direct).
Other		

Assessor (Students, Faculty, Program Leaders, Peer Reviewer, Others (specify))

Assessment Methods (Direct, Indirect)

G. Specification Approval Data

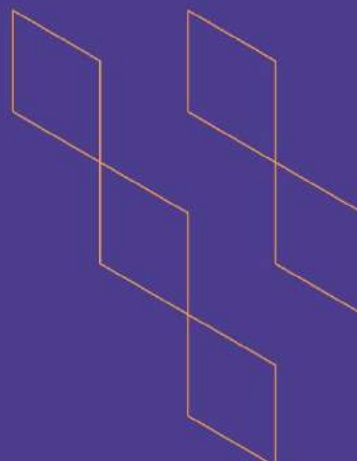
COUNCIL /COMMITTEE	VETERINARY DEPARTMENT
REFERENCE NO.	1444-10-95
DATE	19.02.2023





T-104
2022

Course Specification



Course Title: Rabbit Diseases.
Course Code: VMD 459.
Program: Bachelor of Veterinary Medicine.
Department: Veterinary Medicine.
College: Agriculture and Veterinary Medicine.
Institution: Qassim University .
Version: T-104 (2022)
Last Revision Date: 1/2/2021 .





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G. Specification Approval Data	6



A. General information about the course:

Course Identification	
1. Credit hours:	2 (1+ 1)
2. Course type	
a.	University <input type="checkbox"/> College <input type="checkbox"/> Department <input checked="" type="checkbox"/> Track <input type="checkbox"/> Others <input type="checkbox"/>
b.	Required <input type="checkbox"/> Elective <input checked="" type="checkbox"/>
3. Level/year at which this course is offered:	
4. Course general Description: This course is concerned with the study with the common diseases of rabbit. The clinical signs, pathological findings and diagnostic tools for rabbit diseases are also included. The course will explain the common diseases affecting rabbits in production as well as in experimental laboratory. Furthermore, the course will intend also with the diagnostic tools of the common diseases and differential diagnosis	
5. Pre-requirements for this course (if any): Anatomy of Domestic Animals, Veterinary pharmacology, General veterinary Pathology and Physics.	
6. Co- requirements for this course (if any): VMD 471, VMD 342	
7. Course Main Objective(s):	
1. The students will acquire the necessary skills of the principles of pathology, diagnosis, prevention and control of rabbit diseases.	
2. Students will be able to analyze data from different resources related the clinical, pathological and diagnostic criteria of the viral diseases affecting rabbits.	
3. The students will study the clinical, pathological and diagnostic criteria of the bacterial, parasitic, nutritional and miscellaneous and fungal diseases affecting rabbits	

1. Teaching mode (mark all that apply)

No	Mode of Instruction	Contact Hours	Percentage
1.	Traditional classroom	45	100%
2.	E-learning		
3.	Hybrid <ul style="list-style-type: none"> • Traditional classroom • E-learning 		
4.	Distance learning		

2. Contact Hours (based on the academic semester)

No	Activity	Contact Hours
1.	Lectures	30
2.	Laboratory/Studio	15
3.	Field	-
4.	Tutorial	-
5.	Others (specify)	-





Total

45

B. Course Learning Outcomes (CLOs), Teaching Strategies and Assessment Methods

Co de	Course Learning Outcomes	Code of CLOs aligned with program	Teaching Strategies	Assessment Methods
1.0	Knowledge and understanding			
1.1	Students will be able to list and recognize knowledge related to diseases of rabbit	K1	<ul style="list-style-type: none"> •Lectures with PowerPoint and movie content on diseases of rabbits •Practical lessons on necropsy and gross lesions of rabbit diseases •Practical lesions on histopathological picture of various diseases of rabbit •External visits for farms and slaughter plants 	<ol style="list-style-type: none"> 1. Quarterly exam 2. Practical exam 3. Final exam
2.0	Skills			
2.2				
3.0	Values, autonomy, and responsibility			
3.1	The students be able to take a decision by working as a team to solve problems related to diagnosis of rabbit diseases.	V1	<ul style="list-style-type: none"> •Students are asked to work in small groups within the course, so they can cooperate and share their skills. • Students will be trained on necropsy of rabbits and writing report and suggestive diagnosis. 	<ul style="list-style-type: none"> •Final written exam with different manners of questions •Practical exam •Evaluation of a written report (Assignments)
3.3	Students should be able to make independent assessment and correct diagnosis of rabbit diseases.	V3	<ul style="list-style-type: none"> •Writing reports about field rabbit farm and exterminate lab visits and methods for diagnosis of rabbit diseases. •Individual presentation. 	<ul style="list-style-type: none"> •Evaluation of a written report (Assignments)

C. Course Content

No	List of Topics	Contact Hours
1.	Introduction on rabbit morphology, biology and clinical chemistry	10
2.	Viral diseases of rabbit (clinical, pathological and diagnostic criteria): Myxomatosis, Shope fibroma, viral papillomatosis, rabbit poxvirus, rabbit	10



	herpesvirus infection, rabbit coronavirus, rabbit rotavirus, rabbit viral hemorrhagic disease, and rabies virus infection.	
3.	Bacterial diseases of rabbit (clinical, pathological and diagnostic criteria): Pasteurellosis (snuffle), Bordatella bronchiseptica, Stapylococcus aureus (ulcerative pododermatitis), Treponematosis, Pseudomonas aeruginosa (moist dermatitis), Clostridiosis, colibacillosis, Lawsonia intracellularis (proliferative enteropathy), salmonellosis, tularemia, Yersiniosis, listeriosis and pyogenic pathogens infection.	15
4.	Mycotic diseases of rabbit (clinical, pathological and diagnostic criteria): Superficial mycosis (Ringworm) and deep and systemic mycosis (Aspergillosis)	15
5.	Nutritional and metabolic diseases of rabbit (clinical, pathological and diagnostic criteria): Vitamin E deficiency, hypervitaminosis Vitamin D, Hypo and hypervitaminosis A, Carbohydrate overload, Pregnancy toxemia, chronic fluorosis	10
6.	Neoplastic/proliferative diseases of rabbit (clinical, pathological and diagnostic criteria)	15
7.	Congenital and miscellaneous diseases of rabbit (clinical, pathological and diagnostic criteria): Malocclusion, Buphthalmia (Congenital Glaucoma), Splay Leg, Endometrial venous aneurysms, Hydrocephalus Vertebral fracture, Trichobezar, Barbering, Ulcerative Pododermatitis Hydrometra, Liver Lobe Torsion, Urolithiasis, Lumbar Hernia	15
Total		90

D. Students Assessment Activities

No	Assessment Activities *	Assessment timing (in week no)	Percentage of Total Assessment Score
1.	First quarterly exam	6	10
2.	Second quarterly exam	12	10
3.	Practical exam	15	5
4.	Final exam	16	50
5.	Participation in the lecture halls	15	5
6.	Field reports	13	5
7.	Team work	13	5
8.	Case study	9	5

*Assessment Activities (i.e., Written test, oral test, oral presentation, group project, essay, etc.)

E. Learning Resources and Facilities

1. References and Learning Resources

Essential References

- Suckow, M. A., Brammer, D. W., Rush, H. G., & Crisp, C. E. (2002). Biology and diseases of rabbits. Laboratory animal medicine, 329.
- Percy, Dean H., and Stephen W. Barthold. Pathology of laboratory rodents and rabbits. John Wiley & Sons, 2013.
- Megan H. Nowland, David W. Brammer, Alexis Garcia, Howard G. Rush, Chapter 10 - Biology and Diseases of Rabbits, Editor(s): James G. Fox, Lynn C. Anderson, Glen M. Otto, Kathleen R. Pritchett-Corning, Mark T. Whary,: In American College of Laboratory Animal Medicine, Laboratory Animal Medicine (Third Edition), Academic Press, 2015, Pages 411-461, ISBN 9780124095274, <https://doi.org/10.1016/B978-0->



	12-409527-4.00010-9.
Supportive References	
Electronic Materials	-www. Google.com -www.youtube.com -www.freemedicalvideos.com
Other Learning Materials	computer-based programs/CD, professional standards or regulations and software. NA

2. Required Facilities and equipment

Items	Resources
facilities (Classrooms, laboratories, exhibition rooms, simulation rooms, etc.)	Air conditioned classroom of a minimum of 35 seats and powered by multimedia equipment. Equipped Veterinary clinic or Veterinary Teaching Hospital.
Technology equipment (projector, smart board, software)	Data show connected to a computer (desktop or laptop), access to the world wide web.
Other equipment (depending on the nature of the specialty)	Slide photographing unit (triheaded microscope – digital camera – monitor). - Color printer for glossy paper A3/A4 for the pathology Studio.

F. Assessment of Course Quality

Assessment Areas/Issues	Assessor	Assessment Methods
Effectiveness of teaching	Students – graduates- faculty – program administration – committee for quality assurance and accreditation	Students survey and questionnaire annual reports prepared by the program administrative committee
Effectiveness of students assessment	Students- Graduates- employers - committee for quality assurance and accreditation	Survey for students – graduates – employers – bi-annual reports of the quality assurance unit
Quality of learning resources	Students – graduates – peer reviewer	Surveys and periodical visits of internal and external per reviewers
The extent to which CLOs have been achieved	Students Instructor	Course Evaluation Survey (Indirect). Checking students' performance in the test (Direct).
Other		

Assessor (Students, Faculty, Program Leaders, Peer Reviewer, Others (specify))

Assessment Methods (Direct, Indirect)

G. Specification Approval Data

COUNCIL /COMMITTEE	VETERINARY DEPARTMENT
REFERENCE NO.	1444-10-95





DATE

19.02.2023



T-104
2022

Course Specification

Course Title: Clinical Pharmacology
Course Code: VMD 464
Program: Bachelor of Veterinary Medicine
Department: Veterinary Medicine
College: Agricultural and Veterinary Medicine
Institution: Qassim University
Version: T-104 (2022)
Last Revision Date: 1-2-2022

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1. References and Learning Resources	5
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G. Specification Approval Data	7

A. General information about the course:

Course Identification	
1. Credit hours:	2 (1+1)
2. Course type	
a.	University <input type="checkbox"/> College <input type="checkbox"/> Department <input type="checkbox"/> Track <input type="checkbox"/> Others <input type="checkbox"/>
b.	Required <input type="checkbox"/> Elective <input checked="" type="checkbox"/>
3. Level/year at which this course is offered:	Level 8 / Fourth year
4. Course general Description	
- Preparation of the student to develop an understanding of the principles of individualization of drug therapy, apply the principles of clinical pharmacology and rational pharmacotherapy in clinical practice and develop skills and attitudes needed to recognize and avoid irrational prescribing.	
5. Pre-requirements for this course (if any): VMD 461- VMD 481	
6. Co- requirements for this course (if any): None	
7. Course Main Objective(s)	
- Preparation of the student to	
- - Understand the main theories related to drug action and pharmacokinetics as well as to know the effect of the particular drug on main body organs as well as therapeutic uses. Understand the basis of drug use in treatment of diseases. Memorize the most recent drugs and evaluate their therapeutic potential. Describe the main side effects and overdose toxicity of drugs. Show the hazards of drug residues on the health and choose the drugs with no withdrawal time.	



1. Teaching mode (mark all that apply)

No	Mode of Instruction	Contact Hours	Percentage
1.	Traditional classroom	60	100%
2.	E-learning		
3.	Hybrid <ul style="list-style-type: none"> • Traditional classroom • E-learning 		
4.	Distance learning		

2. Contact Hours (based on the academic semester)

No	Activity	Contact Hours
1.	Lectures	30
2.	Laboratory/Studio	30
3.	Field	
4.	Tutorial	
5.	Others (specify)	
	Total	60

B. Course Learning Outcomes (CLOs), Teaching Strategies and Assessment Methods

Code	Course Learning Outcomes	Code of CLOs aligned with program	Teaching Strategies	Assessment Methods
1.0	Knowledge and understanding			
1.1	The student should be able to understand principles of individualization of drug therapy.	K1	-Lectures. -Practical and clinical sessions. -Take home assignments.	- Examinations, quizzes, clinical procedure evaluated.
1.2				
...				
2.0	Skills			
2.3	The student should be able to apply the principles of clinical pharmacology and rational pharmacotherapy	S3	-Take home: the student, given problems that need to be solved. - Comprehensiv	-Take home: Seminars need to be prepared, presented and will be evaluated



Code	Course Learning Outcomes	Code of CLOs aligned with program	Teaching Strategies	Assessment Methods
	ity in clinical practice and develop skills and attitudes needed to recognize and avoid irrational prescribing		e clinical case , presented to the student and will be asked to formulate a	accordingly -Clinical reports related question discussed in the format of group discussion.
2.4	The student should be able to construct therapeutic plan to a given health problem based on proper evaluation of drug action, kinetics and side effects.	S4		-Students evaluated for their professional attitude by monitoring students throughout their study period.
2.5	The student should be able to clinical simulation of the animal owner.	S5	- Demonstration in computer. -Group assignment. -Using the internet -Clinical simulation of the animal owner. - Communicatio n with animal owners of different levels of educations	-Students assignments evaluated by direct assessment and continuous assessments. -Direct evaluation of take home work.
3.0	Values, autonomy, and responsibility			
3.1				



C. Course Content

No	List of Topics	Contact Hours
1.	Orientation to the course	4
2.	Principles of pharmacokinetics and rational drug therapy	4
3.	Drugs of food and nonfood animals.	4
4.	Drug-drug interactions	4
5.	Potential toxicity of major drug groups in animals.	4
6.	Drugs prohibited for use in food animals	4
7.	Drugs acting on skin, eye and inflammations	4
8.	Growth promoting agents	4
9.	Prudent usage of antimicrobials	4
10.	Fluid therapy	4
11.	Bioavailability	4
12.	Bioequivalence	4
13.	Immunotherapy	4
14.	Gene therapy	4
15.	Veterinary herbal medicine	4
Total		60

D. Students Assessment Activities

No	Assessment Activities *	Assessment timing (in week no)	Percentage of Total Assessment Score
1.	1st midterm	6	10
2.	Assignments and quizzes	4, 8	10
3.	2nd midterm	11	10
4.	Practical exam	15	20
5.	Final Exam	16	50

*Assessment Activities (i.e., Written test, oral test, oral presentation, group project, essay, etc.)

E. Learning Resources and Facilities

1. References and Learning Resources

Essential References	- Goodman L.S. & Gilman A. (2009): The Pharmacological Basis of Therapeutics. 8th Ed. Bailliere Tindal, London. - Harvey, R.A. and Champ P. C. (2009): Lippincott's Illustrated Reviews Pharmacology. 4th Ed. Lippincott Williams and Wilkins.
Supportive References	-Handouts prepared especially for this course, translated from the appropriate textbooks in the field. These handouts are available at the Faculty photocopying center.



	- Basic & Clinical Pharmacology & Toxicology. -Journal of Veterinary Pharmacology and Therapeutics. -Journal of Environmental Toxicology and Pharmacology. -Journal of Pharmacology and Experimental Therapeutics. - Research in Vet. Sci. - Toxicology. Appl. Pharmacology.
Electronic Materials	www.vmd.gov.uk www.usp.org www.fda.gov/search.html www.osha.gov. http://dx.doi.org/10.1007/978-0-387-72573-4 http://dx.doi.org/10.1007/b138634 http://dx.doi.org/10.1007/b138972 http://dx.doi.org/10.1007/978-3-540-38918-7 http://dx.doi.org/10.1007/978-3-540-74805-2 http://dx.doi.org/10.1007/3-7643-7440-3
Other Learning Materials	Suitable videos and animations from the websites.

2. Required Facilities and equipment

Items	Resources
facilities (Classrooms, laboratories, exhibition rooms, simulation rooms, etc.)	Classroom of 20 students capacity. Laboratory of 15students capacity.
Technology equipment (projector, smart board, software)	Data show.
Other equipment (depending on the nature of the specialty)	Chemicals. Glasses and kits for experimental sessions.

F. Assessment of Course Quality

Assessment Areas/Issues	Assessor	Assessment Methods
Effectiveness of teaching	Students	Course Evaluation Survey (Indirect)
Effectiveness of students assessment	<ul style="list-style-type: none"> Students Program Leaders 	<ul style="list-style-type: none"> Course Evaluation Survey (Indirect). Result of the course (Direct)
Quality of learning resources	Students	Course Evaluation Survey (Indirect)
The extent to which CLOs have been achieved	<ul style="list-style-type: none"> Students Instructor 	<ul style="list-style-type: none"> Course Evaluation Survey (Indirect). Checking students' performance in the test (Direct).
Other		



Assessor (Students, Faculty, Program Leaders, Peer Reviewer, Others (specify))

Assessment Methods (Direct, Indirect)

G. Specification Approval Data

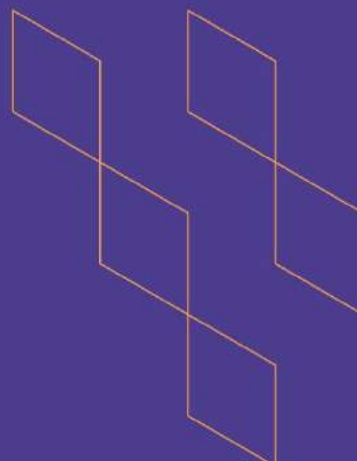
COUNCIL /COMMITTEE	COUNCIL OF VETERINARY MEDICINE DEPARTMENT
REFERENCE NO.	1444-10-95
DATE	19/2/2023





T-104
2022

Course Specification



Course Title:	safety Poultry Meat
Course Code:	478 VMD
Program:	Veterinary Medicine
Department:	Veterinary Medicine
College:	Agriculture and Veterinary Medicine
Institution:	Qassim University
Version:	T-104
Last Revision Date:	1/2/2021 (modified on 10/8/2022 to fit the three semesters system).



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2. Required Facilities and Equipment	8
F. Assessment of Course Quality	8
G. Specification Approval Data	9





A. General information about the course:

Course Identification

1. Credit hours: 2 (1 + 1)

2. Course type

a. University College Department Track Others

b. Required Elective

3. Level/year at which this course is offered:

Level 8/4th year

4. Course general Description

This course discusses the safety aspects of poultry meat production from farm to table. The major chapters of this course include poultry-slaughtering process; sanitation and safety; foodborne pathogens in poultry meat production; HACCP; pre- and post- harvest interventions.

5. Pre-requirements for this course (if any): 344 VMD/ 352 VMD

6. Co- requirements for this course (if any): None

7. Course Main Objective(s)

- A. To gain a basic understanding of the poultry-slaughtering process.
- B. Enabling students to control and prevent the risk of poultry contamination with different hazards.
- C. Enabling students to identify the major sources of poultry contamination.
- D. Enabling students to evaluate the safety of poultry products.
- E. Students will be able to understand and adhere to HACCP regulations throughout the harvest process.

1. Teaching mode (mark all that apply)

No	Mode of Instruction	Contact Hours	Percentage
1.	Traditional classroom	√	100%
2.	E-learning		
	Hybrid		
3.	<ul style="list-style-type: none"> • Traditional classroom • E-learning 		
4.	Distance learning		

2. Contact Hours (based on the academic semester)

No	Activity	Contact Hours
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1.	Lectures	12
2.	Laboratory/Studio	20
3.	Field	3
4.	Tutorial	
5.	Others (specify)	
	Total	35



B. Course Learning Outcomes (CLOs), Teaching Strategies and Assessment Methods

Code	Course Learning Outcomes	Code of CLOs aligned with program	Teaching Strategies	Assessment Methods
1.0	Knowledge and understanding			
1.1	Students will gain a basic understanding of poultry-slaughtering process.	K1	-Lectures -Practical sessions -Field visits	1. Midterm exam 2. Final exam 3. Quizzes 4. Class Participation
2.0	Skills			
2.1	Students will be able to monitor the production of poultry meat to prevent public health hazards.	S3	-Lectures -Practical sessions -Field visits	1. Midterm exam 2. Practical exam 3. Final exam 4. Quizzes 5. Class Participation
2.2	Students will be able to demonstrate analyses of research findings relevant to poultry hygiene.	S5	- Using scholarly sources. - Writing reports. - Searching peer-reviewed journals for information; Presentation of a study's findings.	1. Midterm exam 2. Practical exam 3. Final exam 4. Quizzes 5. Reports
3.0	Values, autonomy, and responsibility			
3.1	Students should be able to make a decision as a team to solve problems related to diagnosis of food poisoning associated with the consumption of poultry	V1	- Writing reports - Group discussion	1. Midterm exam 2. Practical exam 3. Final exam 4. Quizzes



Code	Course Learning Outcomes	Code of CLOs aligned with program	Teaching Strategies	Assessment Methods
				5. Reports
3.2	Students will be able to make an independent assessment of the safety of poultry products.	V3	- Writing reports -Field visits - Practical sessions	1. Midterm exam 2. Practical exam 3. Final exam 4. Quizzes 5. Reports

C. Course Content

No	List of Topics	Contact Hours
1	Introduction	1
2	Poultry-slaughtering process: Premortem handling	1
3	Poultry-slaughtering process: Postmortem handling	1
4	Sanitation and safety: Chemical contaminations	1
5	Sanitation and safety: Microbial contaminations	1
6	Salmonella in Poultry Meat Production	1
7	Antibiotic-Resistant Salmonella	1
8	Campylobacter in Poultry Meat Production	1
9	HACCP	1
10	Pre-harvest Interventions to Improve Poultry Meat Safety	1
11	Post-harvest Interventions to Improve Poultry Meat Safety	1
12	Inspection	1
Total		12

D. Students Assessment Activities

No	Assessment Activities *	Assessment timing (in week no)	Percentage of Total Assessment Score
1	Midterm exam.	6	20%
2	Practical exam.	12	15%
3	Quizzes	Throughout the semester	5%
4	Individual report	12	5%





No	Assessment Activities *	Assessment timing (in week no)	Percentage of Total Assessment Score
5	Class participation	Throughout the semester	5%
6	Final exam.	13	50%

*Assessment Activities (i.e., Written test, oral test, oral presentation, group project, essay, etc.)



E. Learning Resources and Facilities

1. References and Learning Resources

Essential References	Owens, Casey M. Poultry meat processing. CRC Press, 2010. Mead, G., ed. Poultry meat processing and quality. Woodhead Publishing, 2004. Venkitanarayanan, Kumar, Siddhartha Thakur, and Steven C. Ricke, eds. Food safety in poultry meat production. Cham, Switzerland: Springer International Publishing, 2019.
Supportive References	Hui, Y. H. Handbook of Poultry Science and Technology, Primary Processing. Vol. 1. John Wiley & Sons, 2010.
Electronic Materials	Saudi FDA, WHO websites
Other Learning Materials	

2. Required Facilities and equipment

Items	Resources
facilities (Classrooms, laboratories, exhibition rooms, simulation rooms, etc.)	Separate laboratory of at least 25 seats
Technology equipment (projector, smart board, software)	Data show Laptop
Other equipment (depending on the nature of the specialty)	

F. Assessment of Course Quality

Assessment Areas/Issues	Assessor	Assessment Methods
Effectiveness of teaching	Students	Course Evaluation Survey (Indirect)
Effectiveness of students assessment	Students; Program Leaders	Course Evaluation Survey (Indirect); Result of the course (Direct)
Quality of learning resources	Students	Course Evaluation Survey (Indirect)
The extent to which CLOs have been achieved	Students; Instructor	Course Evaluation Survey (Indirect). Checking students' performance in the test (Direct).
Other		

Assessor (Students, Faculty, Program Leaders, Peer Reviewer, Others (specify))

Assessment Methods (Direct, Indirect)





G. Specification Approval Data

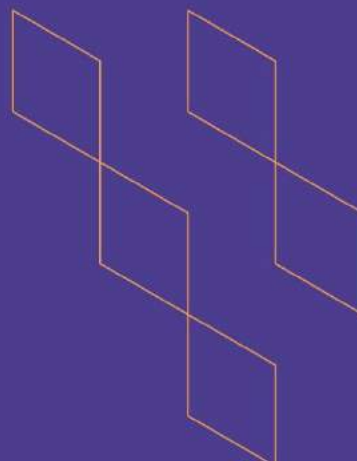
COUNCIL /COMMITTEE	Veterinary Department
REFERENCE NO.	
DATE	





T-104

Course Specification



Course Title: Organic animal production
Course Code: APP 480
Program: Animal production
Department: Animal production and breeding
College: Agriculture and veterinary medicine
Institution: Qassim university
Version: Course Specification Version Number
Last Revision Date: Feb 2023



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A. General information about the course:

Course Identification	
1. Credit hours:	2(1+1)
2. Course type	
a.	University <input type="checkbox"/> College <input checked="" type="checkbox"/> Department <input type="checkbox"/> Track <input type="checkbox"/> Others <input type="checkbox"/>
b.	Required <input type="checkbox"/> Elective <input checked="" type="checkbox"/>
3. Level/year at which this course is offered:6-8 level	
4. Course general Description	
1- Principles and practices of organic farming	
2- The differences between raising traditional and organic farm animals	
3- learning marketing of organic products	
5. Pre-requirements for this course (if any):	
101 Zool	
6. Co- requirements for this course (if any):	
N/A	
7. Course Main Objective(s)	
Raising the knowledge and practices of the students in the field of livestock organic farming	

1. Teaching mode (mark all that apply)

No	Mode of Instruction	Contact Hours	Percentage
1.	Traditional classroom	45	100
2.	E-learning		
3.	Hybrid <ul style="list-style-type: none"> • Traditional classroom • E-learning 		
4.	Distance learning		





2. Contact Hours (based on the academic semester)

No	Activity	Contact Hours
1.	Lectures	30
2.	Laboratory/Studio	
3.	Field	
4.	Tutorial	
5.	Others (specify)	15
	Total	45



B. Course Learning Outcomes (CLOs), Teaching Strategies and Assessment Methods

Code	Course Learning Outcomes	Code of CLOs aligned with program	Teaching Strategies	Assessment Methods
1.0	Knowledge and understanding			
1.1	The student will be able to implement the principles of organic animal farming knowledge and practices		Lectures, Practical sessions, visits to organic animal farms.	Periodical and practical tests. Final exam Quizzes
1.2				
...				
2.0	Skills			
2.1	To be able to choose suitable solutions for the problems related to Organic animal breeding.		Lectures	Quizzes
2.2				
...				
3.0	Values, autonomy, and responsibility			
3.1	Introducing the student to the ethics of dealing and caring for the product from organic animals		lectures	periodic and final exams
3.2				
...				

C. Course Content

No	List of Topics	Contact Hours
1.	Principles and practices of organic farming	6
2.	Farms as ecosystems	3



3.	Certification process and agencies	3
4.	Managing organic matter to support soil nutrients	3
5.	Biodiversity management	3
6.	Forage crop cycles	3
7.	Plant competition and ground cover	3
8.	Differences between conventional and organic farm animal farming	6
9.	Integration of crops and animals	3
10.	Organic animal husbandry practices	6
11.	Shelter for birds and animals	3
12.	Marketing of organic animal products	3
Total		45

D. Students Assessment Activities

No	Assessment Activities *	Assessment timing (in week no)	Percentage of Total Assessment Score
1.	Principles and practices of organic farming	1-3	12%
2.	Farms as ecosystems	4-5	8%
3.	Certification process and agencies	5-6	8%
4.	Managing organic matter to support soil nutrients	6-7	8%
5.	Biodiversity management	7-8	8%
6.	Forage crop cycles	8-9	8%
7.	Plant competition and ground cover	9-10	8%
8.	Differences between conventional and organic farm animal farming	10-11	8%
9.	Integration of crops and animals	11-12	8%
10.	Organic animal husbandry practices	12-13	8%
11.	Shelter for birds and animals	13-14	8%
1	Marketing of organic animal products	14-15	8%
2.			

*Assessment Activities (i.e., Written test, oral test, oral presentation, group project, essay, etc.)





E. Learning Resources and Facilities

1. References and Learning Resources

Essential References	The complete guide to organic livestock farming. Terri Paajanen. (Atlantic Publishing Group 2011). (صالح شعبان عبد الرحمن: الدواجن العضوية) وزارة الزراعة السعودية 3311 دليل الزراعة العضوية (وزارة الزراعة السعودية 3313
Supportive References	1-J. Organic Agriculture 2- Korean J. Org. Agric. 3-International J. Organic farming
Electronic Materials	https://www.infonet-biovision.org/AnimalHealth/Organic-animal-husbandry-Breeding-housing-and-feeding-animals
Other Learning Materials	N/A

2. Required Facilities and equipment

Items	Resources
facilities (Classrooms, laboratories, exhibition rooms, simulation rooms, etc.)	
Technology equipment (projector, smart board, software)	
Other equipment (depending on the nature of the specialty)	


F. Assessment of Course Quality

Assessment Areas/Issues	Assessor	Assessment Methods
Effectiveness of teaching		
Effectiveness of students assessment		
Quality of learning resources		
The extent to which CLOs have been achieved		
Other		

Assessor (Students, Faculty, Program Leaders, Peer Reviewer, Others (specify))

Assessment Methods (Direct, Indirect)

G. Specification Approval Data

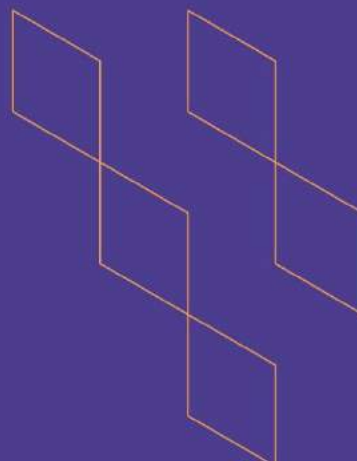
COUNCIL /COMMITTEE	The report has been revised and approved by the departmental council.	
REFERENCE NO.		
DATE	20/3/2023	





T-104
2022

Course Specification



Course Title: Pet Animals Medicine
Course Code: VMD 493
Program: Bachelor of Veterinary Medicine
Department: Veterinary Medicine
College: Agriculture and Veterinary Medicine
Institution: Qassim University
Version: T-104 (2022)
Last Revision Date: 1/2/2021.



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F. Assessment of Course Quality	5
G. Specification Approval Data	6



A. General information about the course:

Course Identification	
1. Credit hours:	2 (1+ 1)
2. Course type	
a.	University <input type="checkbox"/> College <input type="checkbox"/> Department <input checked="" type="checkbox"/> Track <input type="checkbox"/> Others <input type="checkbox"/>
b.	Required <input type="checkbox"/> Elective <input checked="" type="checkbox"/>
3. Level/year at which this course is offered: Elective	
4. Course general Description: This course is concerned with the study of diagnosis and treatment of different diseases of pet animals such as diseases of digestive, respiratory, urogenital system. Veterinary ophthalmology and liver and skin disorders.	
5. Pre-requirements for this course (if any): Virology (VMD 344), Systemic Pathology (VMD 352).	
6. Co- requirements for this course (if any): NA	
7. Course Main Objective(s): After completion of this course, students are expected to know; normal gait of dogs and cats and diagnosis of lameness. Students are expected to know diseases of digestive system, respiratory system, urogenital system. Veterinary ophthalmology and liver and skin disorders.	

1. Teaching mode (mark all that apply)

No	Mode of Instruction	Contact Hours	Percentage
1.	Traditional classroom	70	70%
2.	E-learning		
3.	Hybrid <ul style="list-style-type: none"> • Traditional classroom • E-learning 		
4.	Distance learning		

2. Contact Hours (based on the academic semester)

No	Activity	Contact Hours
1.	Lectures	70
2.	Laboratory/Studio	-
3.	Field	-
4.	Tutorial	-
5.	Others (specify)	30
	Total	100





B. Course Learning Outcomes (CLOs), Teaching Strategies and Assessment Methods

Co de	Course Learning Outcomes	Code of CLOs aligned with program	Teaching Strategies	Assessment Methods
1.0	Knowledge and understanding			
1.1				
2.0	Skills			
2.2	Students will be able to apply critical thinking and analytical skills for intervention in pet animal medicine.	S2	Lectures contain case studies, pictures of clinical cases, and examples of diseases. Case studies are major parts of the practical part of this course. These case studies are upon clinical cases, how to diagnose and methods of treatment.	1. Quarterly exam 2. Practical exam 3. Final exam
3.0	Values, autonomy, and responsibility			
3.1	Students will be able to work effectively in teams for solving medical disorders	V1	Students are trained to work in team. This team is responsible for a clinical case. Each student has a specific task at this team and should perform his task efficiently.	7. Team work
3.2	Students will be able to show full awareness of ethical and professional issues relevant to medical diseases in pets	V2		
3.3	Students will be able to manipulate veterinary field works using sense and locomotor organs.	V3		

C. Course Content

No	List of Topics	Contact Hours
1.	Manipulation of pet animals	12
2.	Diseases of the cardiovascular system of pet animals	12
3.	Diseases of the respiratory system of pet animals	12
4.	Diseases of the digestive system of pet animals	6
5.	Diseases of the urinary system of pet animals	12
6.	Diseases of the skin of pet animals	6
7.	Ocular affections of pet animals	6
8.	Liver disorders of pet animals	6
9.	Malnutrition diseases of pet animals	12
Total		84





D. Students Assessment Activities

No	Assessment Activities *	Assessment timing (in week no)	Percentage of Total Assessment Score
1.	First quarterly exam	6	10
2.	Second quarterly exam	12	10
3.	Practical exam	15	5
4.	Final exam	16	50
5.	Participation in the lecture halls	15	5
6.	Field reports	13	5
7.	Team work	13	5
8.	Case study	9	5

*Assessment Activities (i.e., Written test, oral test, oral presentation, group project, essay, etc.)

E. Learning Resources and Facilities

1. References and Learning Resources

Essential References	Small Animal Internal Medicine 6th Edition by Richard W. Nelson DVM, C. Guillermo Couto DVM. 2019. Practical small animal ultrasonography. Abdomen – April 6, 2016 by Panagiotis Mantis. 2016. Quick guidebook to canine and feline ophthalmology - 2nd edition by Javier Esteban Martín, Lori Newman. 2019 Differential Diagnosis in Small Animal Medicine 2nd Edition by Alex Gough, Kathryn F. Murph. 2015. Cardiovascular Disease in Small Animal Medicine 1st Edition, by Wendy Ware. 2011
Supportive References	Journal of Veterinary Internal Medicine
Electronic Materials	NA
Other Learning Materials	computer-based programs/CD, professional standards or regulations and software.

2. Required Facilities and equipment

Items	Resources
facilities (Classrooms, laboratories, exhibition rooms, simulation rooms, etc.)	Air-conditioned classroom of a minimum of 35 seats and powered by multimedia equipment. Equipped Veterinary clinic or Veterinary Teaching Hospital.
Technology equipment (projector, smart board, software)	Data show connected to a computer (desktop or laptop), access to the world wide web.
Other equipment (depending on the nature of the specialty)	Sterilization equipment; boiler, hot air oven. Autoclave, anesthetic machines, X-ray machine and accessories, Sonography and other diagnostic imaging devices.

F. Assessment of Course Quality

Assessment Areas/Issues	Assessor	Assessment Methods
Effectiveness of teaching	Students – graduates- faculty –	Students survey and





Assessment Areas/Issues	Assessor	Assessment Methods
	program administration – committee for quality assurance and accreditation	questionnaire annual reports prepared by the program administrative committee
Effectiveness of students assessment	Students- Graduates- employers - committee for quality assurance and accreditation	Survey for students – graduates – employers – bi-annual reports of the quality assurance unit
Quality of learning resources	Students – graduates – peer reviewer	Surveys and periodical visits of internal and external per reviewers
The extent to which CLOs have been achieved	Students Instructor	Course Evaluation Survey (Indirect). Checking students' performance in the test (Direct).
Other		

Assessor (Students, Faculty, Program Leaders, Peer Reviewer, Others (specify))

Assessment Methods (Direct, Indirect)

G. Specification Approval Data

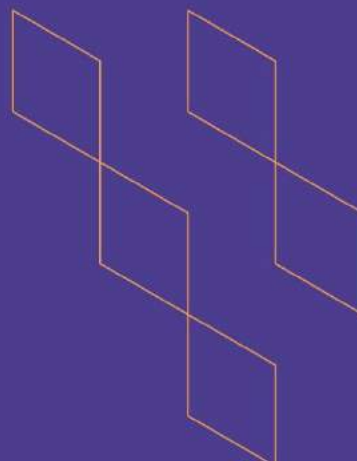
COUNCIL /COMMITTEE	COUNCIL OF VETERINARY MEDICINE DEPARTMENT
REFERENCE NO.	1444-10-95
DATE	19/2/2023





T-104
2022

Course Specification



Course Title: Pet Animals Surgery.
Course Code: VMD 494.
Program: Bachelor of Veterinary Medicine.
Department: Veterinary Medicine.
College: Agriculture and Veterinary Medicine.
Institution: Qassim University .
Version: T-104 (2022)
Last Revision Date: 1/2/2021 .



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F. Assessment of Course Quality	5
G. Specification Approval Data	6



A. General information about the course:

Course Identification	
1. Credit hours:	2 (1+ 1)
2. Course type	
a.	University <input type="checkbox"/> College <input type="checkbox"/> Department <input checked="" type="checkbox"/> Track <input type="checkbox"/> Others <input type="checkbox"/>
b.	Required <input type="checkbox"/> Elective <input checked="" type="checkbox"/>
3. Level/year at which this course is offered: Elective	
4. Course general Description: This course is concerned with the study of diagnosis and treatment of different surgical affections in pet animals such as Gait construction, abnormalities and lameness diagnosis, Affections of the forelimbs, Affections of the hind limbs, Affections of the digestive system, Affections of the respirator system, Affections of the urogenital system, and Veterinary ophthalmology and Veterinary anesthesia.	
5. Pre-requirements for this course (if any): Surgery 1 (VMD 485).	
6. Co- requirements for this course (if any): NA	
7. Course Main Objective(s): After completion of this course, students are expected to know; normal gait of dogs and cats and diagnosis of lameness. Students are expected to know surgical affections of the digestive system, respiratory system, urogenital system. Veterinary ophthalmology and veterinary anesthesia.	

1. Teaching mode (mark all that apply)

No	Mode of Instruction	Contact Hours	Percentage
1.	Traditional classroom	70	70%
2.	E-learning		
3.	Hybrid <ul style="list-style-type: none"> • Traditional classroom • E-learning 		
4.	Distance learning		

2. Contact Hours (based on the academic semester)

No	Activity	Contact Hours
1.	Lectures	70
2.	Laboratory/Studio	-
3.	Field	-
4.	Tutorial	-
5.	Others (specify)	30
	Total	100



B. Course Learning Outcomes (CLOs), Teaching Strategies and Assessment Methods

Co de	Course Learning Outcomes	Code of CLOs aligned with program	Teaching Strategies	Assessment Methods
1.0	Knowledge and understanding			
1.1				
2.0	Skills			
2.2	Students will be able to apply critical thinking and analytical skills for intervention in surgery.	S2	Lectures contain case studies, pictures of clinical cases, and examples of diseases. Case studies are major parts of the practical part of this course. These case studies are upon clinical cases, how to diagnose and methods of treatment.	1. Quarterly exam 2. Practical exam 3. Final exam
3.0	Values, autonomy, and responsibility			
3.1	Students will be able to work effectively in teams for solving veterinary surgical problems.	V1	Students are trained to work in team. This team is responsible for a clinical case. Each student has a specific task at this team and should perform his task efficiently.	7. Team work
3.2	Students will be able to show full awareness of ethical and professional issues relevant to veterinary surgery.	V2		
3.3	Students will be able to manipulate veterinary field works using sense and locomotor organs.	V3	Training and surgery at the practical part of the course	2. Practical exam 11. case study 12. team work

C. Course Content

No	List of Topics	Contact Hours
1.	Gait construction, abnormalities and lameness diagnosis	12
2.	Affections of the forelimbs	12
3.	Affections of the hind limbs	12
4.	Fractures Managements	6
5.	Affections of the digestive system	12
6.	Affections of the respirator system	6
7.	Affections of the urogenital system	6
8.	Common anaesthetic techniques use in pets	6
9.	Veterinary ophthalmology	12
Total		84





D. Students Assessment Activities

No	Assessment Activities *	Assessment timing (in week no)	Percentage of Total Assessment Score
1.	First quarterly exam	6	10
2.	Second quarterly exam	12	10
3.	Practical exam	15	5
4.	Final exam	16	50
5.	Participation in the lecture halls	15	5
6.	Field reports	13	5
7.	Team work	13	5
8.	Case study	9	5

*Assessment Activities (i.e., Written test, oral test, oral presentation, group project, essay, etc.)

E. Learning Resources and Facilities

1. References and Learning Resources

Essential References	Stephen D. Gilson. (2006): Small Animal Soft Tissue Surgery. Clarke KW, Trim, CM, and Hall LW (2014): Veterinary Anaesthesia. Donald Piermattei (2006): Small Animals Orthopedics and Fracture Repair. Douglas Slatter (2003): Small Animal Surgery. Thrall DE (2002): Veterinary diagnostic radiology
Supportive References	Veterinary Surgery Journal
Electronic Materials	WWW contains some video clips on suturing, wounds and fractures, animal anaesthesia.
Other Learning Materials	computer-based programs/CD, professional standards or regulations and software. NA

2. Required Facilities and equipment

Items	Resources
facilities (Classrooms, laboratories, exhibition rooms, simulation rooms, etc.)	Air conditioned classroom of a minimum of 35 seats and powered by multimedia equipment. Equipped Veterinary clinic or Veterinary Teaching Hospital.
Technology equipment (projector, smart board, software)	Data show connected to a computer (desktop or laptop), access to the world wide web.
Other equipment (depending on the nature of the specialty)	Sterilization equipment; boiler, hot air oven. Autoclave, anesthetic machines, X-ray machine and accessories, Sonography and other diagnostic imaging devices. Surgical instruments, sutures and carcasses for training are necessary.

F. Assessment of Course Quality

Assessment Areas/Issues	Assessor	Assessment Methods
Effectiveness of teaching	Students – graduates- faculty – program administration – committee for quality assurance and accreditation	Students survey and questionnaire annual reports prepared by the program administrative



Assessment Areas/Issues	Assessor	Assessment Methods
		committee
Effectiveness of students assessment	Students- Graduates- employers - committee for quality assurance and accreditation	Survey for students – graduates – employers – bi-annual reports of the quality assurance unit
Quality of learning resources	Students – graduates – peer reviewer	Surveys and periodical visits of internal and external per reviewers
The extent to which CLOs have been achieved	Students Instructor	Course Evaluation Survey (Indirect). Checking students' performance in the test (Direct).
Other		

Assessor (Students, Faculty, Program Leaders, Peer Reviewer, Others (specify))

Assessment Methods (Direct, Indirect)

G. Specification Approval Data

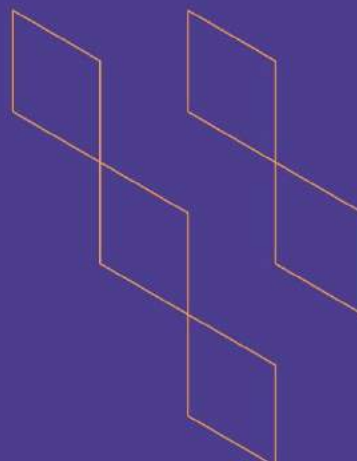
COUNCIL /COMMITTEE	COUNCIL OF VETERINARY MEDICINE DEPARTMENT
REFERENCE NO.	1444-10-95
DATE	19/2/2023





T-104
2022

Course Specification



Course Title: Theriogenology of Pet Animals
Course Code: VMD 495
Program: Bachelor of Veterinary Medicine
Department: Veterinary Medicine
College: Agriculture and Veterinary Medicine
Institution: Qassim University
Version: T-104 (2022)
Last Revision Date: 1/2/2021



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2. Required Facilities and Equipment	6
F. Assessment of Course Quality	6
G. Specification Approval Data	7



A. General information about the course:

Course Identification	
1. Credit hours:	2 (1 + 1)
2. Course type	
a.	University <input type="checkbox"/> College <input type="checkbox"/> Department <input checked="" type="checkbox"/> Track <input type="checkbox"/> Others <input type="checkbox"/>
b.	Required <input type="checkbox"/> Elective <input checked="" type="checkbox"/>
3. Level/year at which this course is offered:	Level Optional
4. Course general Description: The course deals with the functional anatomy of the male and female pets' genital organs, reproductive processes, endocrine control of reproduction, and examination off the reproductive system, reproductive technologies, and infertility in both sexes. The course aims to provide the students with the skills to diagnose and treat different infertility problems in both male and female pet animals.	
5. Pre-requirements for this course (if any): 333 VMD – 342 VMD– 462 VMD	
6. Co- requirements for this course (if any): None	
7. Course Main Objective(s)	
<ul style="list-style-type: none"> The student will be able to diagnose, treat and control reproduction diseases of pet animals The student will be able to diagnose, treat and control of nutritional causes of reproductive diseases in pet animals The student will be able to diagnose infectious causes of reproductive diseases in pet animals. The student will be able to use modern reproductive techniques to improve fertility of pet animals. 	

1. Teaching mode (mark all that apply)

No	Mode of Instruction	Contact Hours	Percentage
1.	Traditional classroom	30	100%
2.	E-learning		
3.	Hybrid <ul style="list-style-type: none"> Traditional classroom E-learning 		
4.	Distance learning		





2. Contact Hours (based on the academic semester)

No	Activity	Contact Hours
1.	Lectures	15
2.	Laboratory/Studio	15
3.	Field	-
4.	Tutorial	-
5.	Others (specify)	-
	Total	30

B. Course Learning Outcomes (CLOs), Teaching Strategies and Assessment Methods

Code	Course Learning Outcomes	Code of CLOs aligned with program	Teaching Strategies	Assessment Methods
1.0	Knowledge and understanding			
2.0	Skills			
2.2	Students will be able to determine comprehensive methods for diagnosis, differential diagnosis, treatment and control of different reproductive disorders of pet animals.	S2	Lectures practical room and clinical cases diagnosis and treatment	2. Practical tests. 4-participation in the lecture hall 13- Oral exam
2.6	Students will be able to manipulate Theriogenology field works using sense and locomotor organs	S6	practical room and clinical cases diagnosis and treatment	1. Quarterly tests. 2. Practical tests. 3. Final tests. 7-field report 11- Case study
3.0	Values, autonomy, and responsibility			
3.1	Students will be able to work effectively in teams for solving infertility problems.	V1	- Lectures - Showing of pictures and Videos of signs and lesions - Case studies	11- Case study 12-group working
3.2	Students will be able to show full awareness of ethical and professional	V2	practical room and clinical cases diagnosis and	2-practical exam 4-participation in the





Code	Course Learning Outcomes	Code of CLOs aligned with program	Teaching Strategies	Assessment Methods
	issues relevant to Veterinary Theriogenology Students will be able to show full awareness of ethical and professional issues relevant to Veterinary Theriogenology		treatment	lecture hall 7-field report 12-group working

C. Course Content

No	List of Topics	Contact Hours
1.	Functional Anatomy of the female pets genital organs	2
2.	Functional Anatomy of the female pets genital organs	2
3.	Estrous cycle, ovulation, and fertilization	2
4.	Estrous cycle, ovulation, and fertilization	2
5.	Examination of non-pregnant and pregnant pets animals	2
6.	Examination of non-pregnant and pregnant pets animals	2
7.	Diagnostic Ultrasonography In pets Reproduction	2
8.	Diagnostic Ultrasonography In pets Reproduction	2
9.	Diseases of the female pets genital organs	2
10.	Diseases of the female pets genital organs	2
11.	Physiology of pregnancy	2
12.	Dystocia in pets	2
13.	Function anatomy of the male pets genital organs	2
14.	Male reproductive diseases	2
15.	Reproductive techniques in pets	2
Total		30

D. Students Assessment Activities

No	Assessment Activities *	Assessment timing (in week no)	Percentage of Total Assessment Score
1	First and second midterm (written test).	7 & 12	20
2	Practical exam	15	5
3	Final exam	16	50
4	Participation in the lecture halls	15	5





No	Assessment Activities *	Assessment timing (in week no)	Percentage of Total Assessment Score
5	Field reports	13	5
6	Team work	13	5
7	Case study	9	5
8	Oral exam	15	5

*Assessment Activities (i.e., Written test, oral test, oral presentation, group project, essay, etc.)

E. Learning Resources and Facilities

1. References and Learning Resources

Essential References	<ol style="list-style-type: none"> 1. Pathways to Pregnancy and Parturition 3rd Edition by P. L. Senger, Current Conceptions Inc., 2012. 2. Holst PA. 1999. Canine Reproduction: The Breeder's Guide 2nd Edition, Alpine Pubns Inc; 2 edition. 3. Margaret V. Root Kustritz: Small Animal Theriogenology: Butterworth-Heinemann, 2003 4. Shirley Dianne Johnston, Margaret V. Root Kustritz, Patricia Schultz Olso. 2001. Canine and Feline Theriogenology, Saunders. 5. Senger PL. 2003. Pathways to Pregnancy and Parturition" 2nd Edition. 6. Morrow DA. 2004. Current Therapy in Theriogenology" W.B. Saunders Company.
Supportive References	Journal of Theriogenology, Animal Reproduction science
Electronic Materials	https://www.arssales.com/ https://visgar.vetmed.ufl.edu/ https://www.nifa.usda.gov/grants/programs/animal-programs/animal-reproduction
Other Learning Materials	Publications related to pet animals reproduction uploaded by the instructors in the research engines as researchgate, Google scholar and science direct.

2. Required Facilities and equipment

Items	Resources
Facilities (Classrooms, laboratories, exhibition rooms,	<ul style="list-style-type: none"> • Classroom of 20 students' capacity. • Laboratory of 15students capacity.



Items	Resources
simulation rooms, etc.)	<ul style="list-style-type: none"> Veterinary hospital with all necessary diagnostic tools and clinical examination
Technology equipment (projector, smart board, software)	<ul style="list-style-type: none"> Data show.
Other equipment (depending on the nature of the specialty)	Ultrasonography, Doppler, diagnostic kits.

F. Assessment of Course Quality

Assessment Areas/Issues	Assessor	Assessment Methods
Effectiveness of teaching	<ul style="list-style-type: none"> Students Committee of Development and Quality Program chairman 	<ul style="list-style-type: none"> Course Evaluation Survey. Frequency of grades distribution form.
Effectiveness of students assessment	<ul style="list-style-type: none"> Committee of Development and Quality Students Program chairman 	<ul style="list-style-type: none"> Internal moderation. Internal benchmarking with similar other course(s). Course Evaluation Survey.
Quality of learning resources	Students	Course Evaluation Survey.
The extent to which CLOs have been achieved	<ul style="list-style-type: none"> Students Instructor 	<ul style="list-style-type: none"> Course Evaluation Survey. Students' performance in the tests and tasks.
The extent to which students are prepared to carry lab. diagnosis and interpret results	Instructor	<ul style="list-style-type: none"> Practical sessions and exams. Assignments.

Assessor (Students, Faculty, Program Leaders, Peer Reviewer, Others (specify))

Assessment Methods (Direct, Indirect)

G. Specification Approval Data

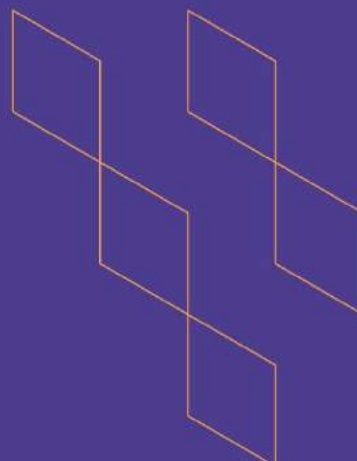
COUNCIL /COMMITTEE	COUNCIL OF VETERINARY MEDICINE DEPARTMENT
REFERENCE NO.	1444-10-95
DATE	19/2/2023





T-104
2022

Course Specification



Course Title: **Wildlife health and management**

Course Code: **VMD 496.**

Program: **Bachelor of Veterinary Medicine.**

Department: **Veterinary Medicine.**

College: **Agriculture and Veterinary Medicine.**

Institution: **Qassim University.**

Version: **T-104 (2022)**

Last Revision Date: **1/2/2021 .**



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A. General information about the course:

Course Identification	
1. Credit hours:	2 (1+ 1)
2. Course type	
a.	University <input type="checkbox"/> College <input checked="" type="checkbox"/> Department <input type="checkbox"/> Track <input type="checkbox"/> Others <input type="checkbox"/>
b.	Required <input type="checkbox"/> Elective <input checked="" type="checkbox"/>
3. Level/year at which this course is offered: any level: from 3rd to 10th levels	
4. Course general Description: This course is concerned with wild animal husbandry and protection including their health and some diseases that they can be infected with. Also, the extinction and the causes of extinction of wild animals, especially the endangered species. the course is covering identify the national regulations and legislation to preserve wildlife and ethical and professional issues relevant to wild animals.	
5. Pre-requirements for this course (if any): NA	
6. Co- requirements for this course (if any): NA	
7. Course Main Objective(s): After completion of the course, students should be able to:	
1. List and recognize knowledge related to wild animal husbandry and protection	
2. Identify the national regulations and legislation to preserve wildlife	
3. Show full awareness of ethical and professional issues relevant to wild animals	
4. Demonstrate strong writing and oral communications skills	

1. Teaching mode (mark all that apply)

No	Mode of Instruction	Contact Hours	Percentage
1.	Traditional classroom	45	90%
2.	E-learning		
3.	Hybrid <ul style="list-style-type: none"> • Traditional classroom • E-learning 		
4.	Distance learning	5	10%

2. Contact Hours (based on the academic semester)

No	Activity	Contact Hours
1.	Lectures	15
2.	Laboratory/Studio	30
3.	Field	-
4.	Tutorial	-
5.	Others (specify)	-
	Total	45





B. Course Learning Outcomes (CLOs), Teaching Strategies and Assessment Methods

Code	Course Learning Outcomes	Code of CLOs aligned with program	Teaching Strategies	Assessment Methods
1.0	Knowledge and understanding			
1.1	Students will be able to list and recognize knowledge related to wild animal husbandry and protection.		1. Lecture 2. Class discussion 3. Homework	1. Participation in classroom 2. Quizzes 3. Midterm-Exam
2.0	Skills			
2.1	Students will be able to determine the national regulations and legislation to preserve wildlife		1. Lecture 2. Class discussion 3. Homework	1. Participation in classroom 2. Quizzes 3. Midterm-Exam 6. Final Exam
2.2	Students will be able to show full awareness of ethical and professional issues relevant to wild animals.		1. Class discussion 2. Homework 3. Showing electronic materials Visiting nature reserves and showing used strategies and systems.	2. Quizzes 3. Midterm-Exam 5. Practical exam 6. Final Exam
3.0	Values, autonomy, and responsibility			
3.1	Students will be able to demonstrate strong writing and oral communications skills.		1. Class discussion 2. Homework	1. Participation in classroom 4. Homework assignments 5. Practical exam
...				

C. Course Content

No	List of Topics	Contact Hours
1.	Introduction of health and the types diseases, source of infection, diagnosis, treatment, and disease control.	7
2.	Some viral diseases that infect wild animals.	7
3.	Some bacterial, parasitic, and fungal diseases that infect wild animals.	8
4.	Wildlife and its values and environmental factors that affect the wildlife.	7
5.	The extinction and the causes of extinction.	8
6.	Husbandry and protection of some wild animals.	8
Total		45





D. Students Assessment Activities

No	Assessment Activities *	Assessment timing (in week no)	Percentage of Total Assessment Score
1.	Class discussion and participation in classroom	Weekly	10
2.	Quiz	4 and 8	6
3.	Midterm-Exam	5 and 11	10
4.	Homework assignments	9	4
5.	Practical exam	14	20
6.	Final Exam	18	50

*Assessment Activities (i.e., Written test, oral test, oral presentation, group project, essay, etc.)

E. Learning Resources and Facilities

1. References and Learning Resources

Essential References	Miller, G.T. and Spoolman, S.E. (2011). Living in the Environment: Principles, Connections, and Solutions Animal Behavior and Wildlife Conservation. Editors: Marco Festa-Bianchet & Marco Apollonio 2003 Saudi Wildlife Authority publications 'in Arabic'
Supportive References	أنظمة المحافظة على الحياة الفطرية والمواطن الطبيعية في المملكة العربية السعودية المها من الأسر إلى التوطين المحميات الطبيعية في المملكة العربية السعودية
Electronic Materials	http://www.swa.gov.sa/ http://www.nwf.org/Wildlife.aspx https://www.cites.org/ http://www.wwf.org/ http://uae.panda.org/ar/ www.endangeredspecies.com www.youtube.com www.alwelaie.com
Other Learning Materials	NA

2. Required Facilities and equipment

Items	Resources
facilities (Classrooms, laboratories, exhibition rooms, simulation rooms, etc.)	Class room for 30 students Well-equipped lab
Technology equipment (projector, smart board, software)	Data show connected to computer (desktop or laptop) Smart board
Other equipment (depending on the nature of the specialty)	Digital camera Video camera





F. Assessment of Course Quality

Assessment Areas/Issues	Assessor	Assessment Methods
Strategies for Obtaining Student Feedback on Effectiveness of Teaching	Students, graduates, faculty, program administration, committee for quality assurance and accreditation	Students survey and questionnaire - annual reports prepared by the program administrative committee
Other Strategies for Evaluation of Teaching by the Program/Department Instructor	Students, graduates, employers, committee for quality assurance and accreditation	Survey for students - graduates - employers – bi-annual reports of the quality assurance unit
Processes for Improvement of Teaching	Students, graduates, and peer reviewer	Surveys and periodical visits of internal and external peer reviewers
Verifying Standards of Student Achievement (e.g., check marking by an independent member teaching staff of a sample of student work, periodic exchange and remarking of tests or a sample of assignments with staff at another institution)	The head of the department revise course grades; maximum, minimum and average. An independent committee appointed by the Head Department.	The program quality assurance and accreditation unit have a committee responsible for revising a sample of courses exams to check marks and submit a report to department head.
Describe the planning arrangements for periodically reviewing course effectiveness and planning for improvement	Students, faculty, and quality unit.	- Students' evaluation form. - Faculty evaluation form. - Periodic checking of students' works - Benchmarking with similar other course(s)
Other		

Assessor (Students, Faculty, Program Leaders, Peer Reviewer, Others (specify))

Assessment Methods (Direct, Indirect)

G. Specification Approval Data

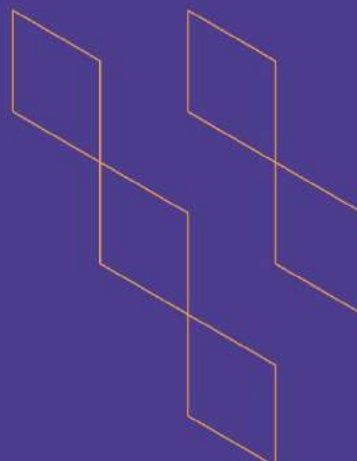
COUNCIL /COMMITTEE	VETERINARY DEPARTMENT
REFERENCE NO.	
DATE	





T-104
2022

Course Specification



Course Title: Wild Animal Anesthesia.
Course Code: VMD 497.
Program: Bachelor of Veterinary Medicine.
Department: Veterinary Medicine.
College: Agriculture and Veterinary Medicine.
Institution: Qassim University .
Version: T-104 (2022)
Last Revision Date: 1/2/2021.



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A. General information about the course:

Course Identification	
1. Credit hours:	2 (1+ 1)
2. Course type	
a.	University <input type="checkbox"/> College <input type="checkbox"/> Department <input checked="" type="checkbox"/> Track <input type="checkbox"/> Others <input type="checkbox"/>
b.	Required <input type="checkbox"/> Elective <input checked="" type="checkbox"/>
3. Level/year at which this course is offered: Elective	
4. Course general Description: This course is concerned with the study of anesthesia and analgesia of wild animals .	
5. Pre-requirements for this course (if any): (VMD 461).	
6. Co- requirements for this course (if any): NA	
7. Course Main Objective(s):	
1. The students will acquire the necessary skills related to sedation, tranquilization, and anesthesia of wild animals.	
2. Students will be able to analyze data from different resources related principles of anesthesia and preparation of wild animals for surgery.	
3. Students will be able to deal with general and special anesthetics instruments, how to handle them practically.	

1. Teaching mode (mark all that apply)

No	Mode of Instruction	Contact Hours	Percentage
1.	Traditional classroom	90	100%
2.	E-learning		
3.	Hybrid <ul style="list-style-type: none"> • Traditional classroom • E-learning 		
4.	Distance learning		

2. Contact Hours (based on the academic semester)

No	Activity	Contact Hours
1.	Lectures	30
2.	Laboratory/Studio	60
3.	Field	-
4.	Tutorial	-
5.	Others (specify)	-
	Total	90





B. Course Learning Outcomes (CLOs), Teaching Strategies and Assessment Methods

Co de	Course Learning Outcomes	Code of CLOs aligned with program	Teaching Strategies	Assessment Methods
1.0	Knowledge and understanding			
1.1				
2.0	Skills			
2.2	Students will be able to apply critical thinking and analytical skills for intervention in Anaesthesia.	S2	Lectures contain case studies, pictures of clinical cases, and examples of diseases. Case studies are major parts of the practical part of this course. These case studies are upon clinical cases, how to deal with anaesthesia.	1. Quarterly exam 2. Practical exam 3. Final exam
2.4	Students will be able to manipulate veterinary field works using sense and locomotor organs.	S6	Training and Anaesthesia at the practical part of the course	2. Practical exam 7. Team work 8. Case study
3.0	Values, autonomy, and responsibility			
3.1	Students will be able to work effectively in teams for solving veterinary Anaesthetic problems.	V1	Students are trained to work in team. This team is responsible for a clinical case. Each student has a specific task at this team and should perform his task efficiently.	7. Team work
3.2	Students will be able to show full awareness of ethical and professional issues relevant to veterinary anaesthesia.	V2		
...				

C. Course Content

No	List of Topics	Contact Hours
1.	Techniques of local Anaesthesia.	10
2.	Anaesthetic agents	10
3.	Techniques of regional anaesthesia	18
4.	Tranquillization	6
5.	Induction of anaesthesia	6
6.	General anaesthesia	6
7.	Anaesthetic machines and accessories	12
8.	Monitoring of anaesthesia	12
9.	Recovery of anaesthesia	10
Total		90





D. Students Assessment Activities

No	Assessment Activities *	Assessment timing (in week no)	Percentage of Total Assessment Score
1.	First quarterly exam	6	10
2.	Second quarterly exam	12	10
3.	Practical exam	15	5
4.	Final exam	16	50
5.	Participation in the lecture halls	15	5
6.	Field reports	13	5
7.	Team work	13	5
8.	Case study	9	5

*Assessment Activities (i.e., Written test, oral test, oral presentation, group project, essay, etc.)

E. Learning Resources and Facilities

1. References and Learning Resources

Essential References	Lumb and Jones' Veterinary Anesthesia and Analgesia, 4th Edition. William J. Tranquilli, John C. Thurmon, Kurt A. Grimm. Wiley-Blackwell. 2007. - Dollar's veterinary surgery general, operative and regional. John A. W. Dollar, James Joseph O'Conner. Baillière, Tindall and Cox, 1930.
Supportive References	Veterinary anaesthesia and analgesia Journal
Electronic Materials	WWW contains some video clips on suturing, wounds and fractures, animal anaesthesia.
Other Learning Materials	computer-based programs/CD, professional standards or regulations and software. NA

2. Required Facilities and equipment

Items	Resources
facilities (Classrooms, laboratories, exhibition rooms, simulation rooms, etc.)	Air conditioned classroom of a minimum of 35 seats and powered by multimedia equipment. Equipped Veterinary clinic or Veterinary Teaching Hospital.
Technology equipment (projector, smart board, software)	Data show connected to a computer (desktop or laptop), access to the world wide web.
Other equipment (depending on the nature of the specialty)	Sterilization equipment; boiler, hot air oven. Autoclaves...etc, and anesthetic machines.

F. Assessment of Course Quality

Assessment Areas/Issues	Assessor	Assessment Methods
Effectiveness of teaching	Students – graduates- faculty – program administration – committee for quality assurance and accreditation	Students survey and questionnaire annual reports prepared by the program administrative committee
Effectiveness of students assessment	Students- Graduates- employers - committee for quality	Survey for students – graduates – employers – bi-





Assessment Areas/Issues	Assessor	Assessment Methods
	assurance and accreditation	annual reports of the quality assurance unit
Quality of learning resources	Students – graduates – peer reviewer	Surveys and periodical visits of internal and external per reviewers
The extent to which CLOs have been achieved	Students Instructor	Course Evaluation Survey (Indirect). Checking students' performance in the test (Direct).
Other		

Assessor (Students, Faculty, Program Leaders, Peer Reviewer, Others (specify))

Assessment Methods (Direct, Indirect)

G. Specification Approval Data

COUNCIL /COMMITTEE	VETERINARY DEPARTMENT
REFERENCE NO.	
DATE	

